

# **Fluorescence Probes and Labels for Biomedical Applications**

**Ewald Terpetschnig**

# Fluorescence Based Biological Assay Formats

*Fluorescence Intensity Assays*

*Fluorescence Polarization Assays*

*Fluorescence Energy Transfer Assays*

*Fluorescence Lifetime Assays*

*Luminescent, Time-Resolved Assays*

# Fluorescence Based Biological Assay Formats

## *Fluorescence Intensity Assays*

- *Most Frequently Used Read-Out Parameter*
- *Prone to Interferences*
- *Not Suitable for Homogeneous Assay*

# Fluorescence Based Biological Assay Formats

## Fluorescence Polarization Assays

*Widely Used for HTS and Clinical Assays*

*Self-Referenced Method*

*Only One Label for Assay Required*

# Fluorescence Based Biological Assay Formats

## Fluorescence Energy Transfer Assays

*Two Labels for Assay Required*

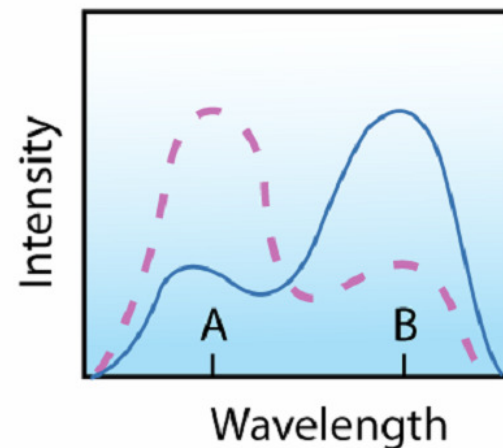
**READ OUT:**

*Intensity (dark acceptor)*

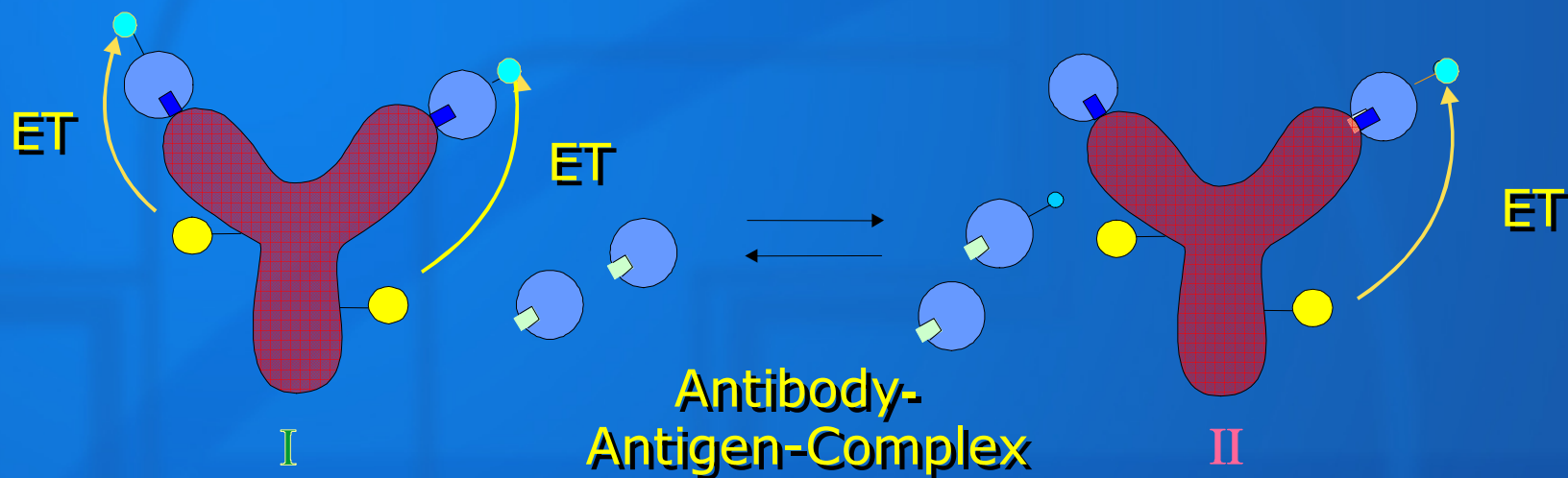
*Intensity-Ratiometric (2 fluorophores)*

*Lifetime (dark acceptor)*

Wavelength-ratiometric method



# Competitive Energy Transfer Immunoassay



- Fluorescent Donor
- Non-fluorescent Acceptor  $\tau(D_{\text{bound}}) < \tau(D_{\text{free}})$
- No Need to Separate D and A Signals
- Lifetime independent of Volume, Color- Quenching, etc

# Fluorescence Based Biological Assay Formats

## *Fluorescence Lifetime Assays*

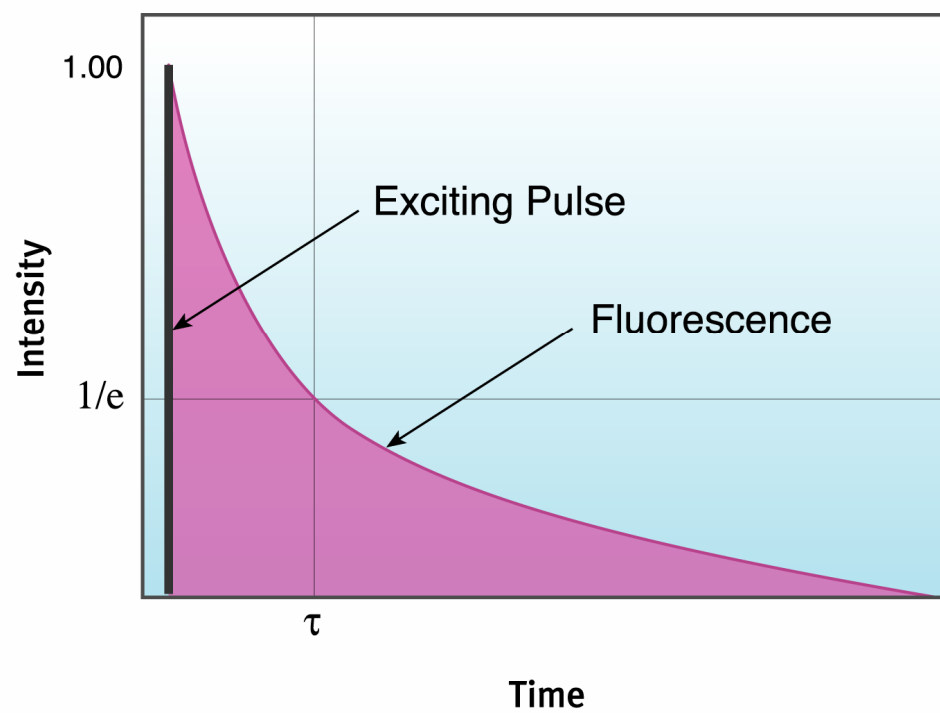
- *Independent of Fluorophore Concentration*
- *1- or 2-Label Assays*

### ***2 Methods:***

- *Phase-Modulation Method*
- *Pulse-Method*

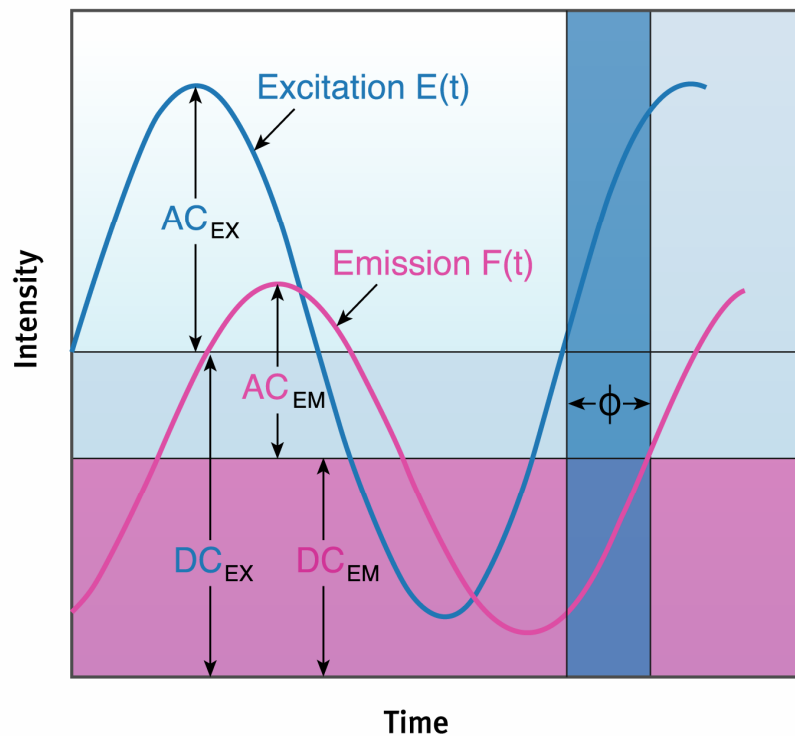
# Two Ways to Measure Lifetime:

# Time-Domain



$$I_t = \alpha e^{-t/\tau}$$

# Frequency Domain



- Phase Shift

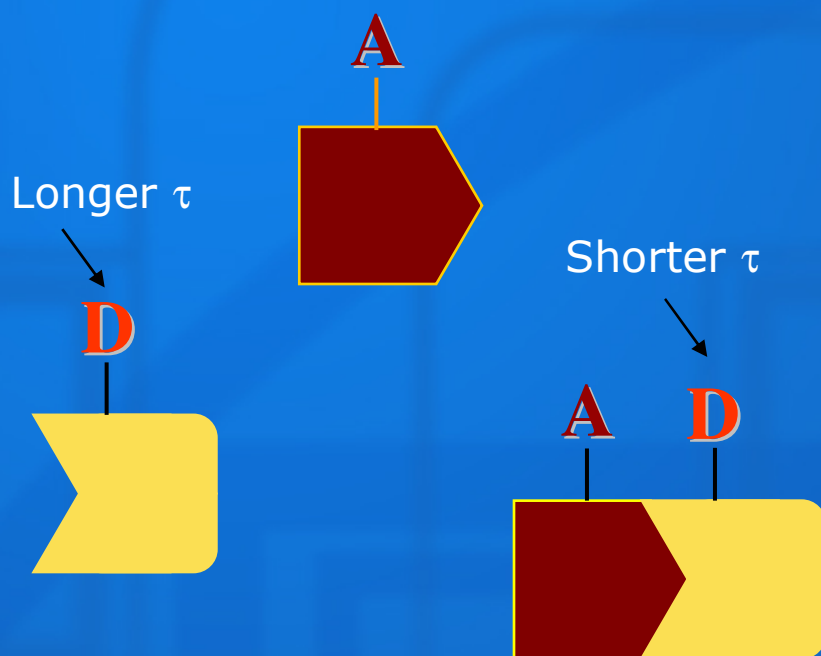
$$\tan \phi = \omega \tau$$

- Demodulation

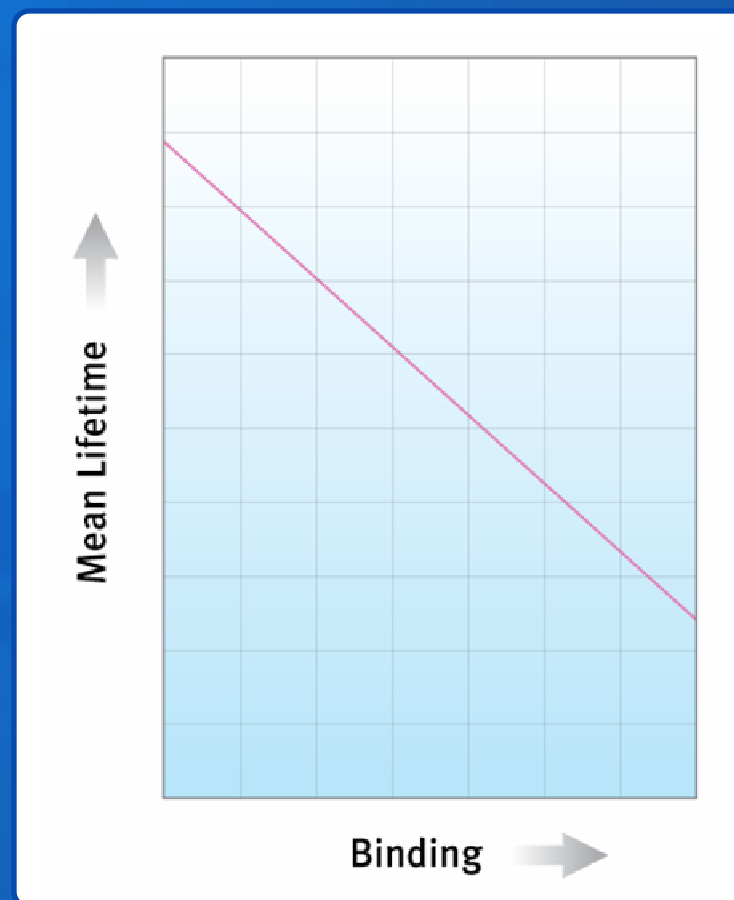
$$M = \frac{(AC/DC)_{EM}}{(AC/DC)_{EX}}$$

$$M = \frac{1}{\sqrt{1 + (\omega \tau)^2}}$$

# Assays Can Be Designed with Lifetime Readout

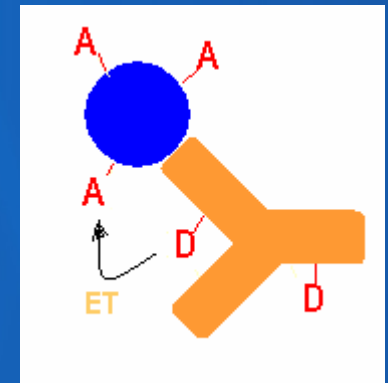


In FRET the Acceptor Shortens the Donor's Lifetime



# Lifetime-Detected Energy Transfer (Binding Assay)

- Fluorescent Donor
- Nonfluorescent Acceptor A
- $\tau (D_{\text{bound}}) < \tau (D_{\text{free}})$
- No need to separate D and A signals
- Lifetime independent of volume, color- quenching, etc.



# Classification:

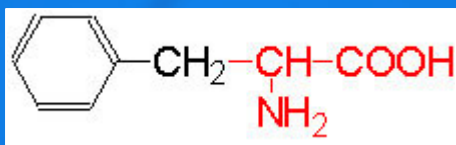
- Intrinsic Fluorophores
- Extrinsic Fluorophores

# Intrinsic Fluorophores

Naturally Occurring Fluorophores

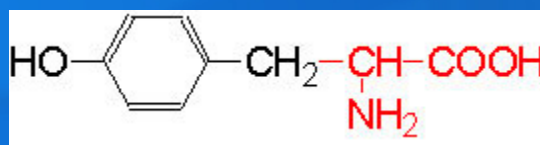
# Proteins: Naturally Occurring Fluorophores

## Aromatic amino acids



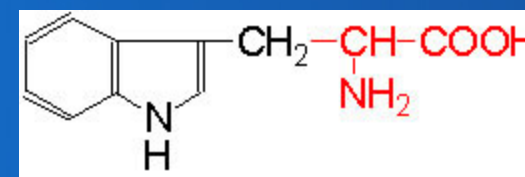
**Phenylalanine (Phe – F)**

Ex/Em 260 nm/282 nm



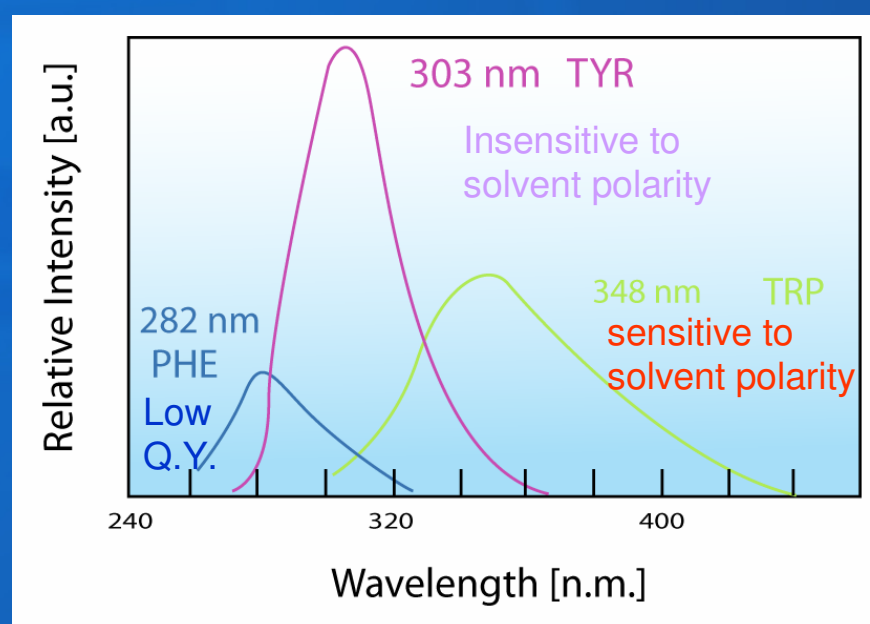
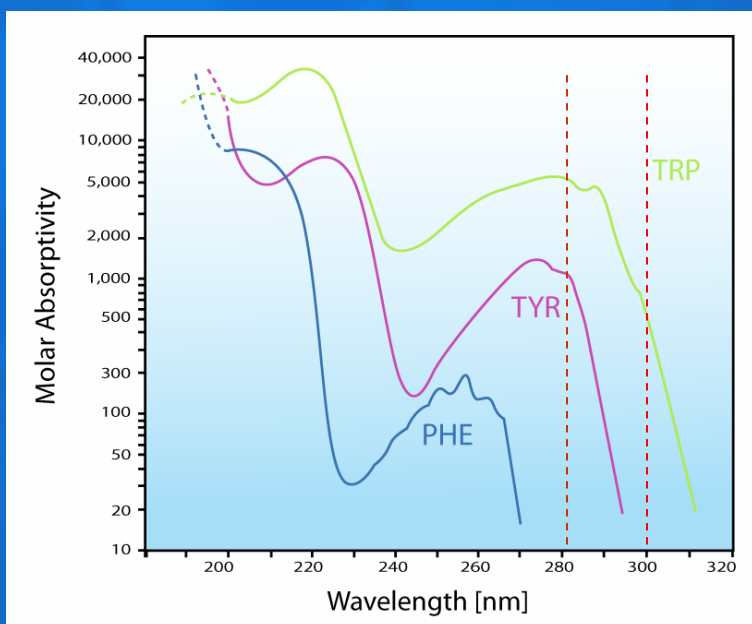
**Tyrosine (Tyr – Y)**

Ex/Em 280 nm/303 nm

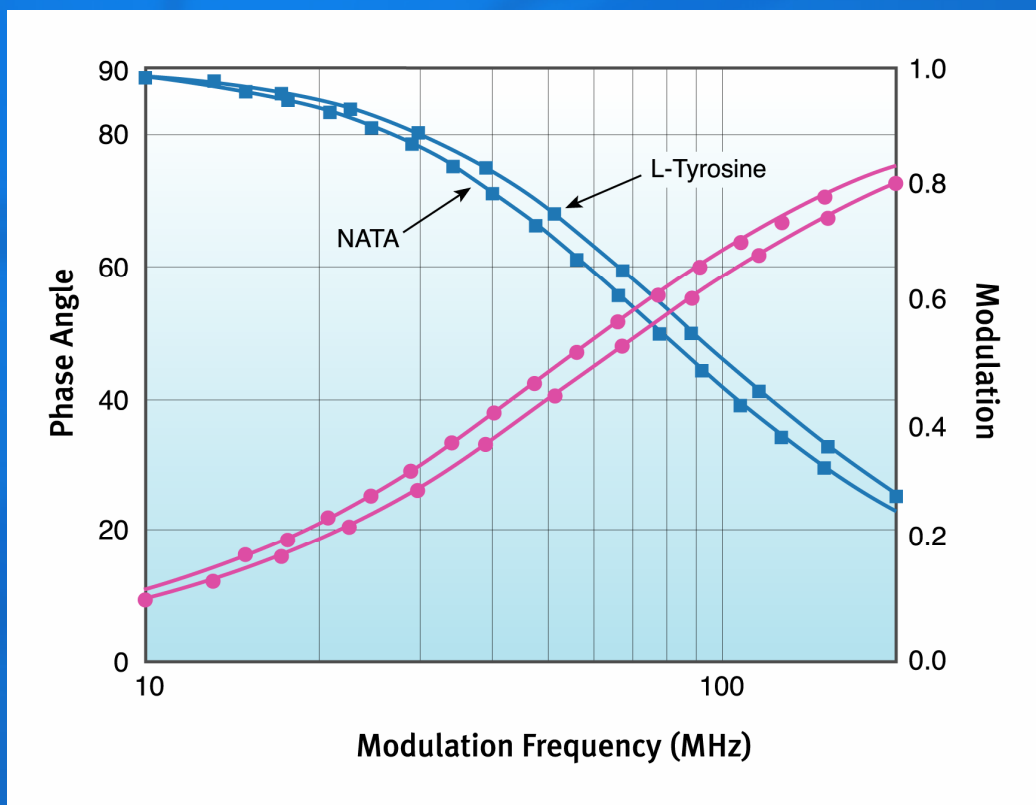


**Tryptophan (Trp-W)**

Ex/Em 280, 295nm/ 305-350 nm



# Fluorescence Lifetimes of Protein-Related Fluorophores



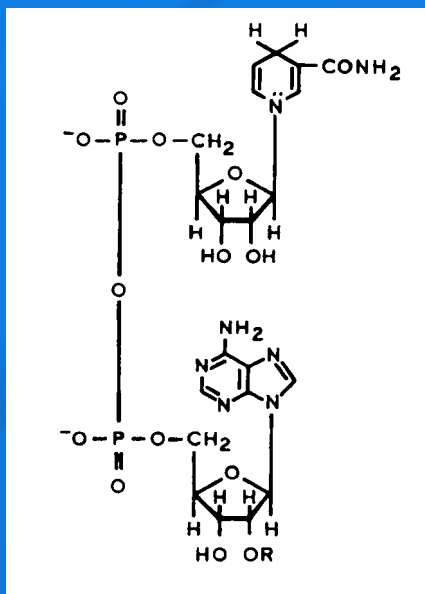
## Tyrosine:

- $\lambda_{\text{Ex}}$ : 280-nm LED  
Em: 320-nm LP
- $\tau = 3.2$  ns (Water)

## NATA:

- $\lambda_{\text{Ex}}$ : 300-nm LED  
Em: 320-nm LP
- $\tau = 3.09$  ns (Water)

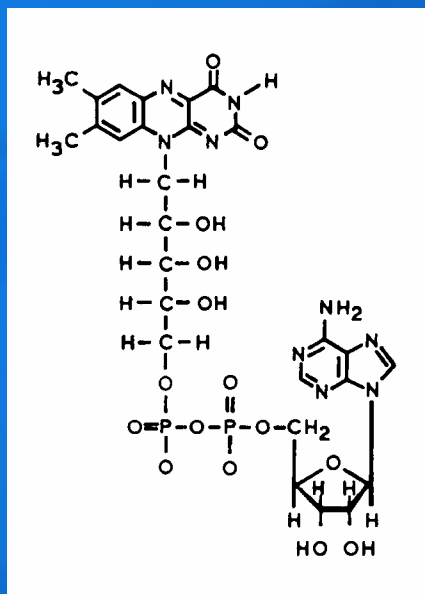
## Enzyme Cofactors



### NADH

(Oxido-reductases)

Ex/Em 340/460 nm



### FAD

(Metabolic enzymes)

Ex/Em 450nm/540 nm

## Porphyrins

Ex/Em 550 nm/620 nm

**Fe<sup>2+</sup> (Heme)**

myoglobin,

hemoglobin

cytochromes b and c,

cytochrome P450 and

cytochrome oxidase

**Mg<sup>2+</sup> chlorophylls**

# Extrinsic Fluorophores

Synthetic dyes or modified biochemicals that are added to a specimen to produce fluorescence with specific spectral properties.

## Fluorescent Probes:

- Non covalent interaction

A fluorescent probe is a fluorophore designed to localize within a specific region of a biological specimen or to respond to a specific analyte.

## Fluorescent Labels:

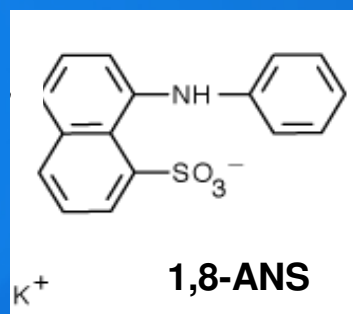
- Covalent interaction

# Classes of Probes, Dyes and Labels:

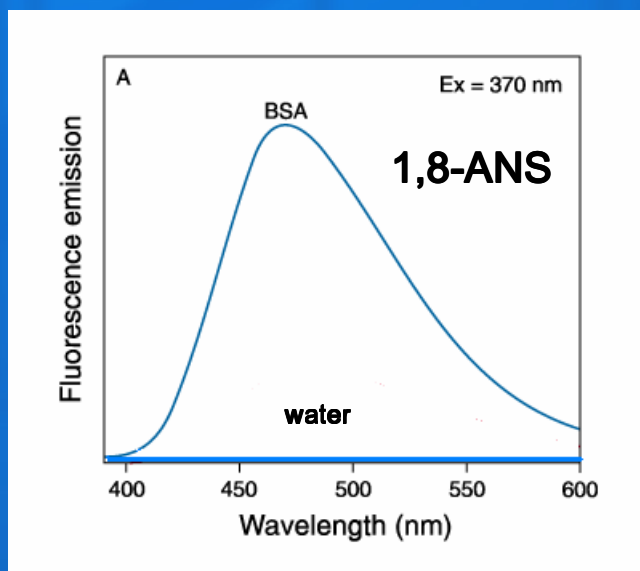
- Organic Dyes
- Metal-Ligand Complexes
- Quantum Dots and Nanoparticles
  - Fluorescent Beads or Polymers
  - Fluorescent Proteins

# Fluorescent Probes

## *Non-covalent*



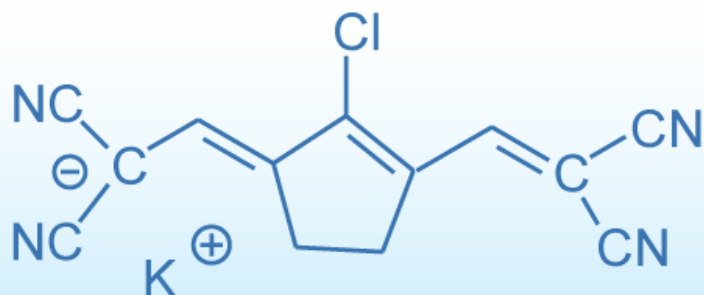
## **1,8-ANS**



Developed by G. Weber in 1950's

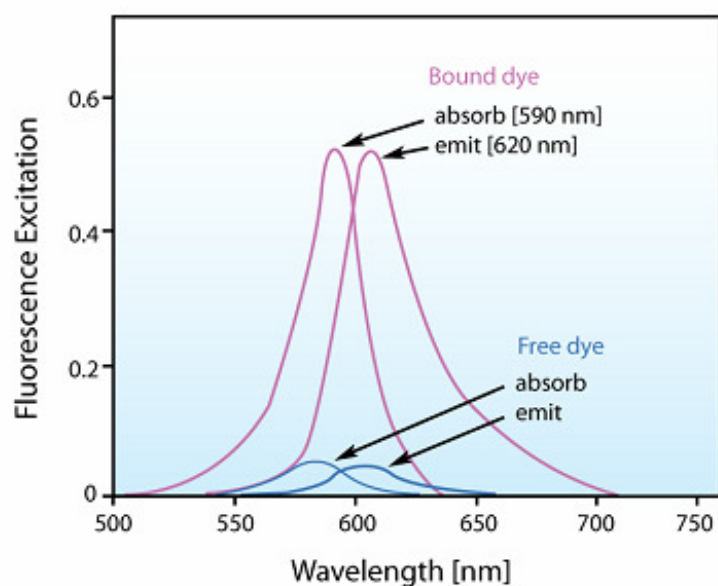
Barely fluorescent water - fluorescence is strongly enhanced in hydrophobic environments

Valuable probes for studying membranes and proteins: QY ~ 0.25 (membranes) or ~0.7 (proteins)



## Albumin Blue

Measurement of albumin levels in biological samples including serum and urine.

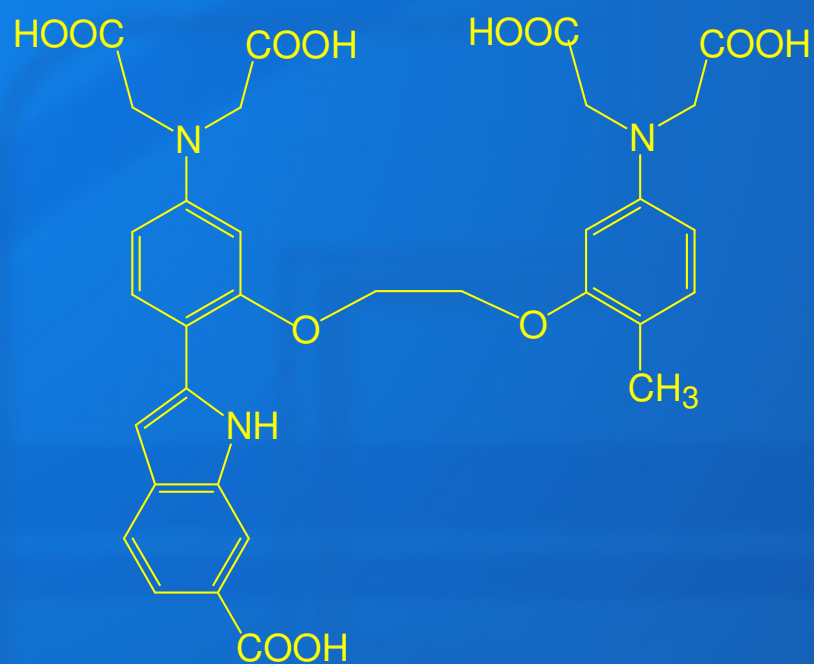


Developed by Kessler & Wolfbeis

Specific- quantitative determination of Albumin in presence of other proteins

Albumin-specific also at concentrations below 100mg.L<sup>-1</sup> - Microalbuminuria

# Fluorescent Ion-Probes



# Fluorescent Ion-Probes

Fluorescence probes have been developed for a wide range of ions:

## Cations:

H<sup>+</sup>, Ca<sup>2+</sup>, Li<sup>+</sup>, Na<sup>+</sup>, K<sup>+</sup>, Mg<sup>2+</sup>, Zn<sup>2+</sup>, Pb<sup>2+</sup> *and others*

## Anions:

Cl<sup>-</sup>, PO<sub>4</sub><sup>2-</sup>, Citrate, ATP, *and others*

# How to choose the correct fluorescent probe

## Dissociation Constant ( $K_d$ )

- Must be compatible with the concentration (pH) range of interest.
- Calibration:  $K_d$  of probe is dependent on pH, temperature, viscosity, ionic strength etc.....

## Measurement Mode

Qualitative or quantitative measurements.

- Ion-probes showing spectral shifts - radiometric measurements
- Light source available

## Indicator Form

Influences cell loading and distribution of the probe.

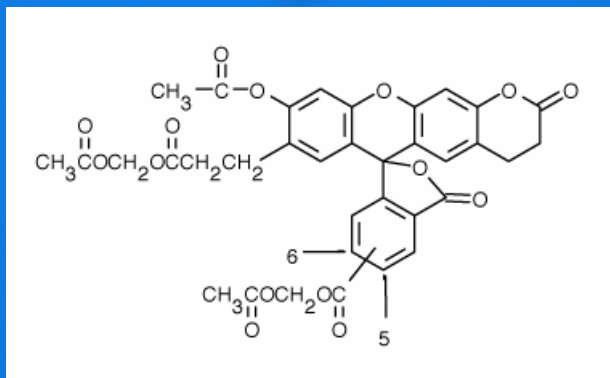
- Salts and dextran-conjugates - microinjection, electroporation
- AM-esters - passively loaded and cleaved by intracellular esterases

# pH-Probes

Probe	pH Range	Measurement Mode
SNARF indicators	6.0-8.0	Em. ratio 580/640 nm
HPTS (pyranine)	7.0-8.0	Exc. ratio 450/405 nm
BCECF	6.5-7.5	Exc. ratio 490/440 nm
Fluoresceins and Carboxyfluoresceins	6.0-7.2	Exc. ratio 490/450 nm
Oregon Green dyes	4.2-5.7	Exc. ratio 510/450 nm
LysoSensor Yellow/Blue DND-160	3.5-6.0	Em. ratio 450/510 nm

Molecular Probes' pH indicator families, in order of decreasing  $pK_a$

# BCECF

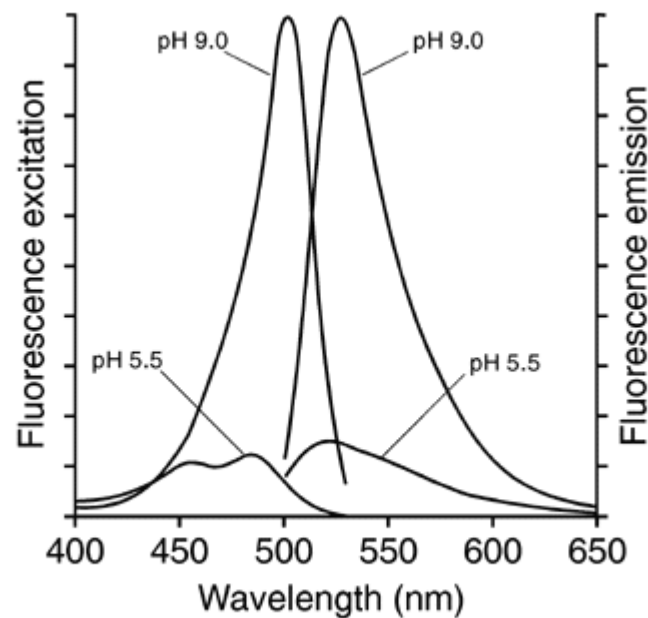


R. Tsien 1982

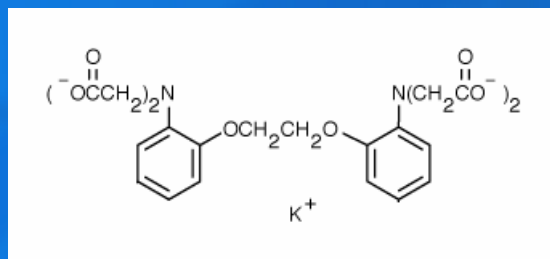
Most widely used fluorescent indicator for intercellular pH

Membrane-permeant AM: pKa ~ 6.98 is ideal for intracellular pH measurements

Excitation-ratiometric probe with  $\lambda_p$  at 439 nm, which is used as the reference point



# Calcium-Probes



## BAPTA

Chelator with high selectivity for  $\text{Ca}^{2+}$  in presence of excess [mM]  $\text{Mg}^{2+}$

**$K_d$  for  $\text{Ca}^{2+}$**

No  $\text{Mg}^{2+}$ : 160 nM

1 mM  $\text{Mg}^{2+}$ : 700 nM

# Calcium-Probes

## UV

**FURA** ( Fura-2, Fura-4F, Fura-5F, Fura-6F, Fura-FF)  
**INDO** ( Indo-1, Indo 5F)

**Ratiometric**

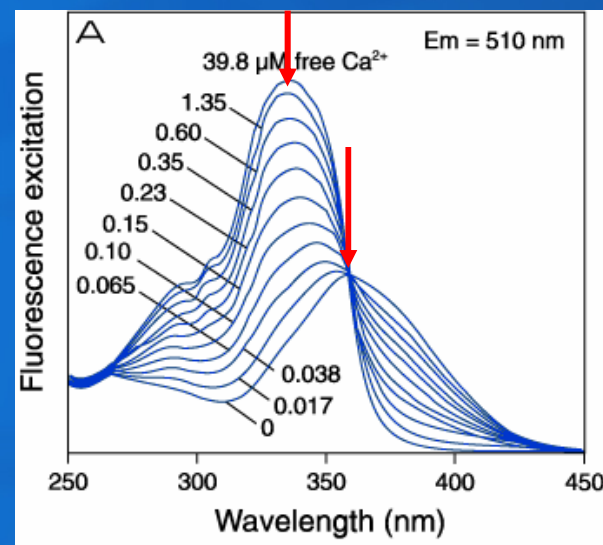
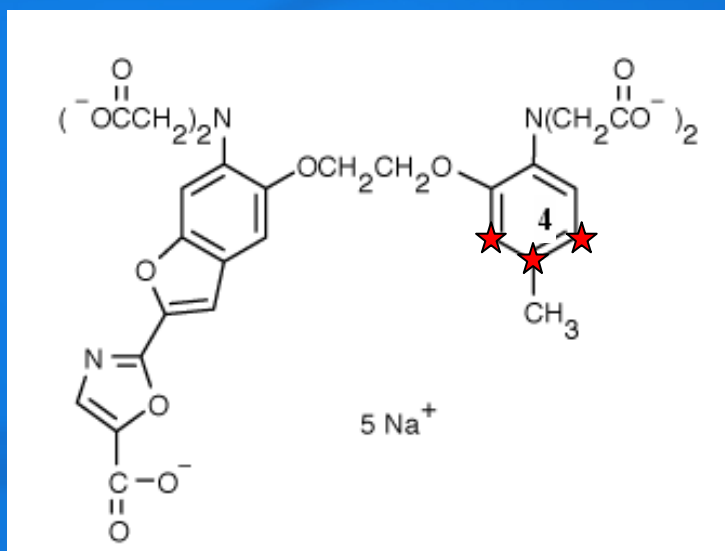
## VISIBLE

**FLUO** (Fluo-3, Fluo-4, Fluo5F, Fluo-5N, Fluo-4N)  
**RHOD** ( Rhod-2, Rhod-FF, Rhod-5N)  
Calcium Green, Calcium Orange, Calcium Crimson  
Oregon Green 488-BAPTA

**Non-  
Ratiometric**

# FURA-2

## Excitation-Ratiometric



Indicator	K <sub>d</sub> (Ca <sup>2+</sup> )
Fura-2	0.14 μM
Fura-5F	0.40 μM
Fura-4F	0.77 μM
Fura-6F	5.30 μM
Fura-FF (5,6)	35 μM

Most used in conventional microscopic imaging

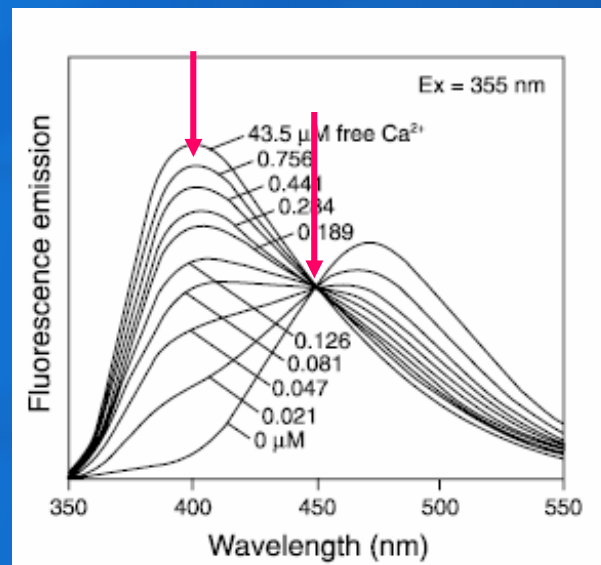
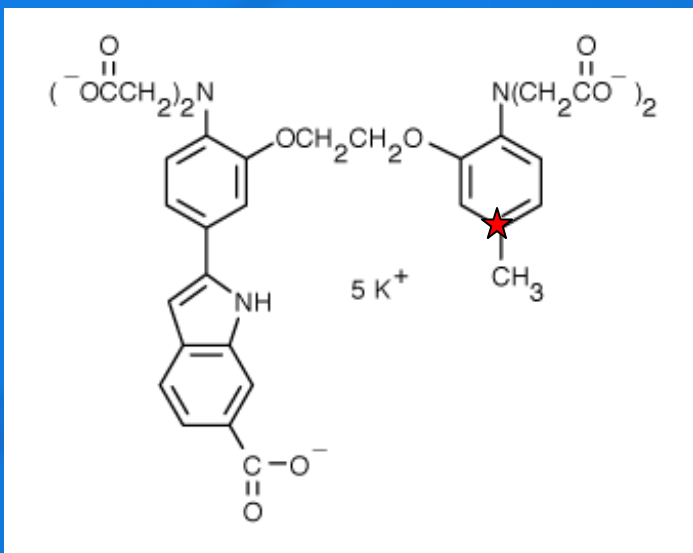
Good excitation shift with Ca<sup>2+</sup>

Ratioed between 340 and 350 and 380 to 385 nm

Fura-4F, Fura-5F, Fura-6F and Fura-FF provide increased sensitivity to intracellular Ca<sup>2+</sup> concentration in the 0.5–35 μM range

# Indo-1

# Emission-Ratiometric



Indicator	K <sub>d</sub> (Ca <sup>2+</sup> ) (μM)
indo-1	0.23
indo-5F	0.47

Most used in laser flow cytometry

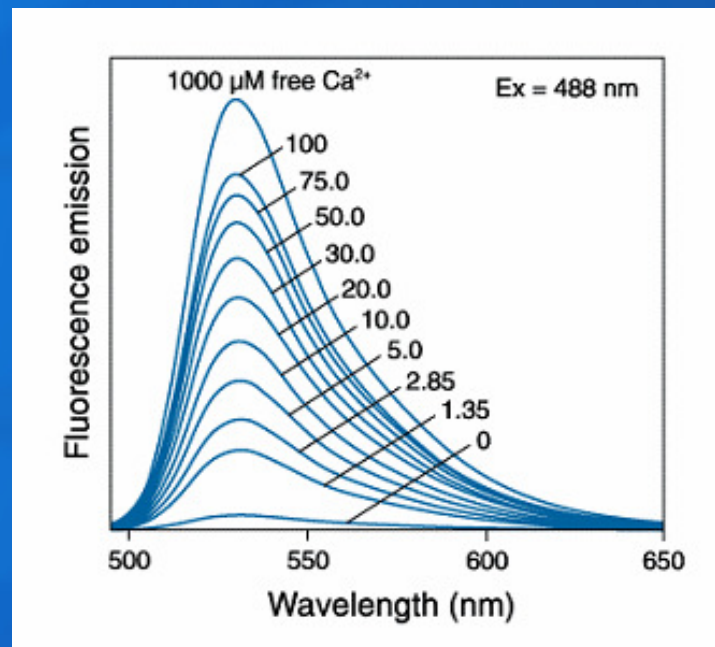
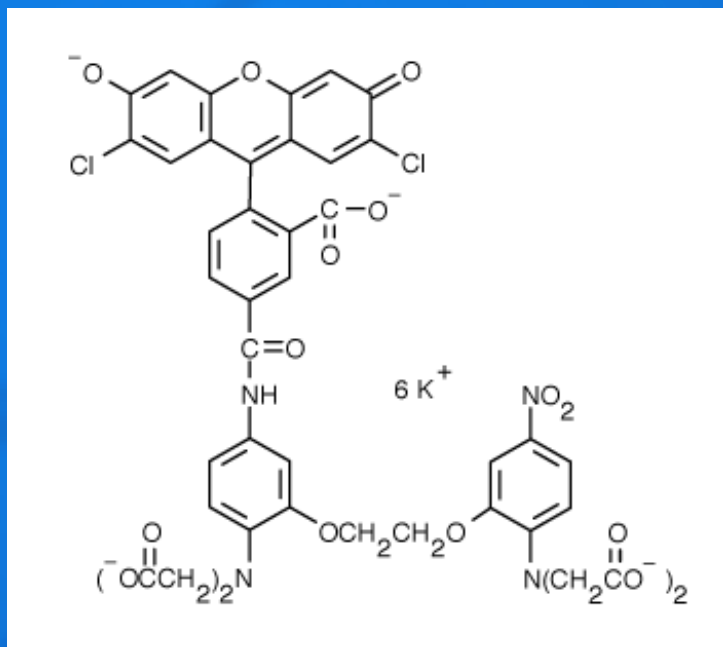
Ratioed between 450 and 405 nm

Photobleaches faster than Fura-2

Excitation with UV laser or Ti-Sapphire at 350 nm

## Calcium Green-5N

## Non-Ratiometric



Indicator	$K_d(\text{Ca}^{2+})$
Calcium Green-5N	14 $\mu\text{M}$

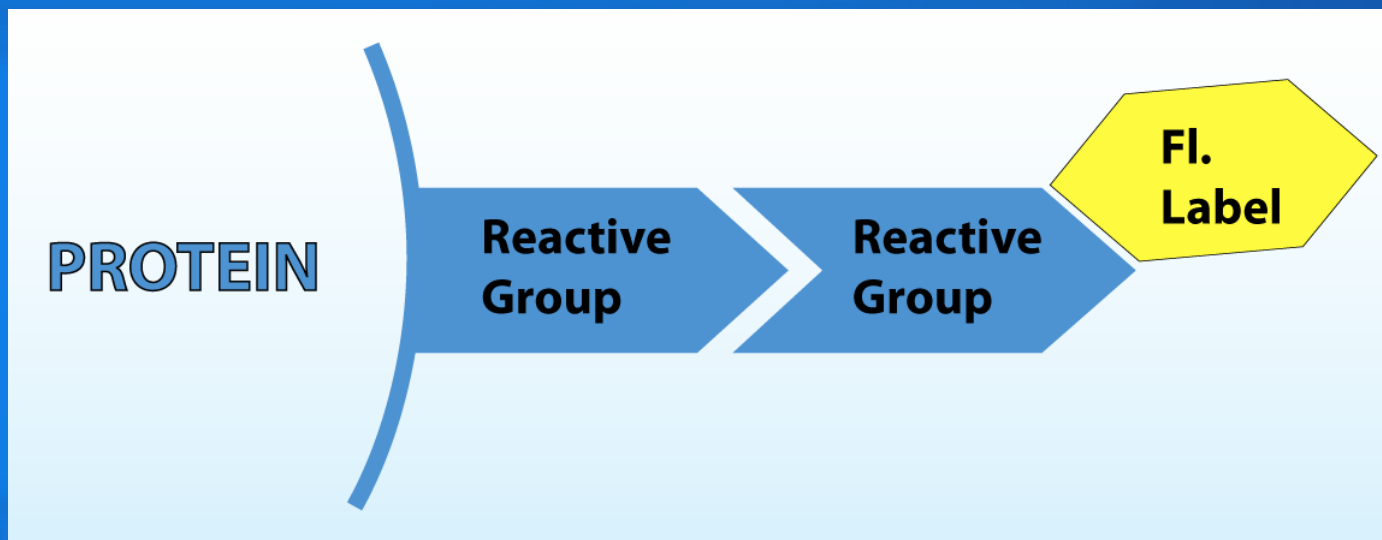
Low affinity Ca-probe

Low fluorescence in absence of  $\text{Ca}^{2+}$

Tracking rapid  $\text{Ca}^{2+}$ -release kinetics



# Selecting the Label



## Reactive groups on proteins

**NH<sub>2</sub>** Lysine  
N-terminus

**SH** Cysteine

Depends on the reactive group on the protein

Light source

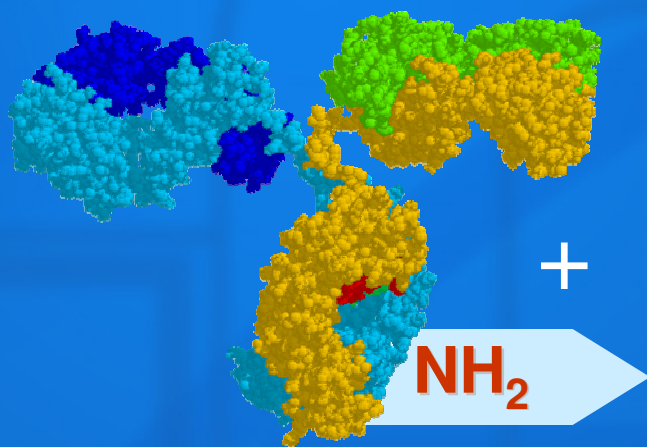
Spectral properties

Autofluorescence

Photostability

**Labeling should not alter the biological activity of biomolecules**

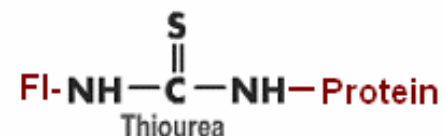
## Amino-Modification:



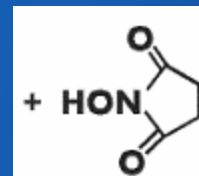
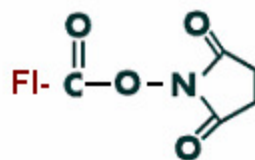
**Protein-NH<sub>2</sub>**

**Lysine  
N-terminus**

isothiocyanate:  
**FI-N=C=S**



succinimidyl ester:



sulfonyl chloride:  
**FI-SO<sub>2</sub>Cl**



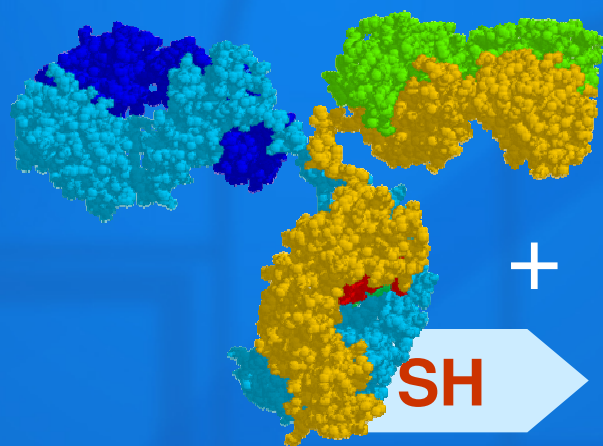
+ HCl

aldehyde:

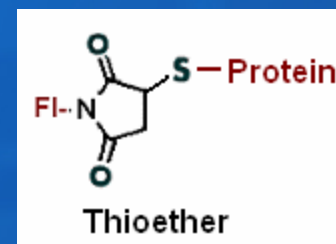
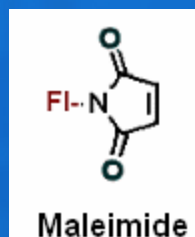
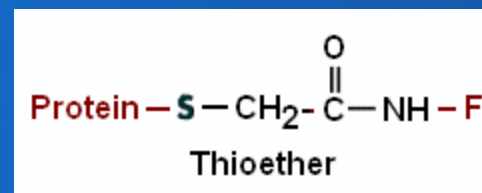
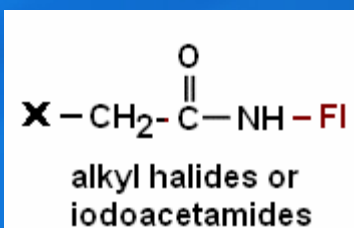


reduction

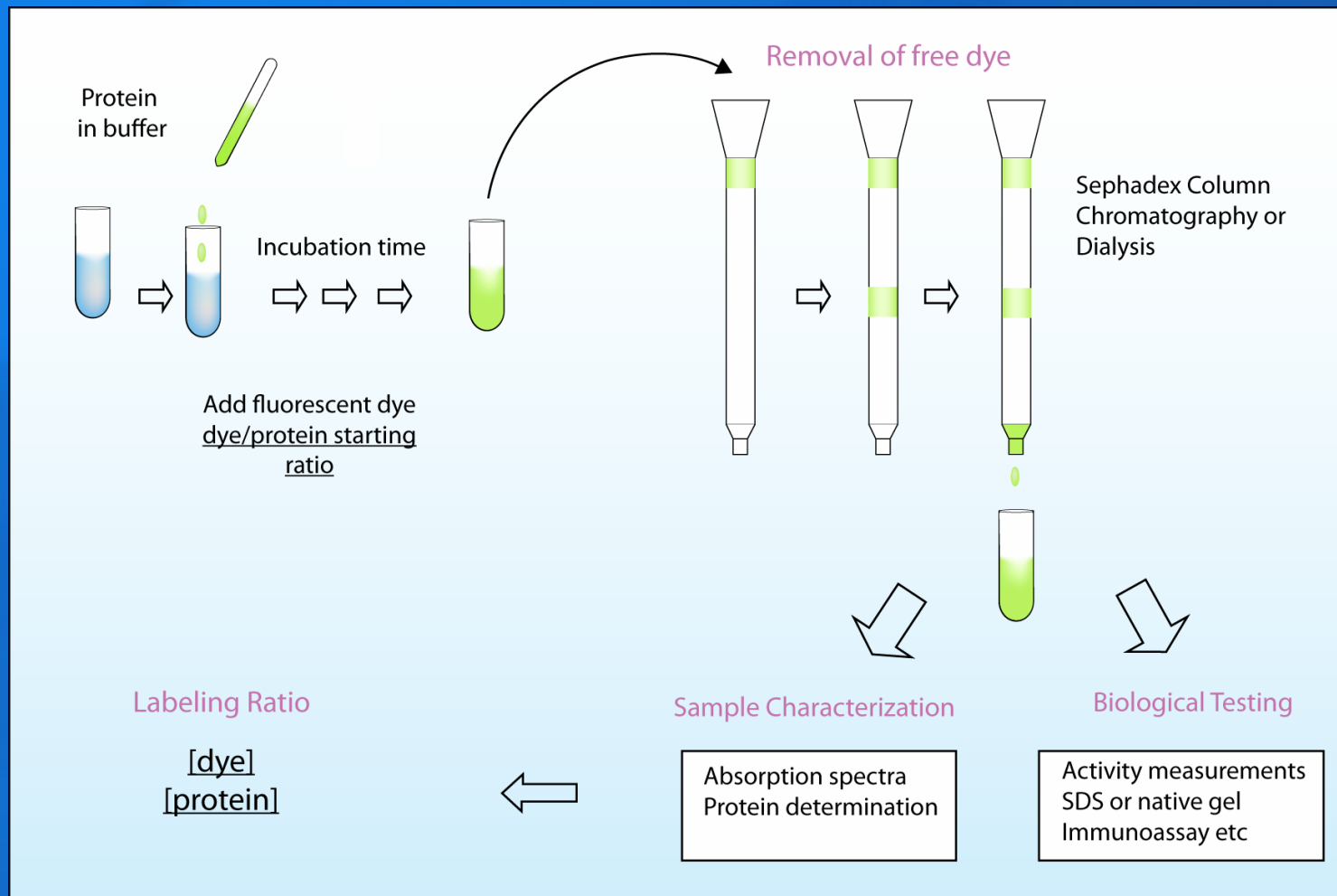
## Thiol-Modification:



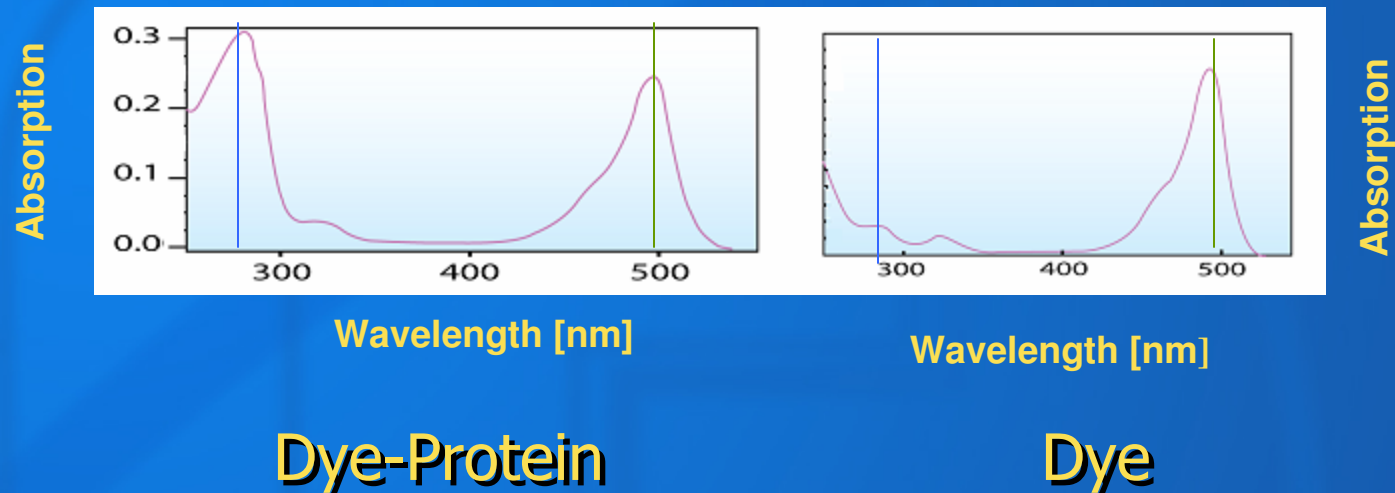
**Protein-SH**  
Cysteine



# Labeling Procedure



# Determination of Dye-to-Protein Ratios



$$D/P = A_{\text{conj}(\lambda_{\text{max}})} \cdot \epsilon_{\text{Prot}} / (A_{\text{conj}(280)} - X \cdot A_{\text{conj}(\lambda_{\text{max}})}) \cdot \epsilon_{\text{dye}}$$

$$X = A_{\text{dye}(280)} / A_{\text{dye}(\lambda_{\text{max}})}$$

Protein:

Photometric measurement

Bradford, Lowry assay

# Organic Dye Classes

## Xanthenes

Fluoresceins - pH-sensitive, not photostable

Rhodamines - tendency to aggregate in aqueous solution

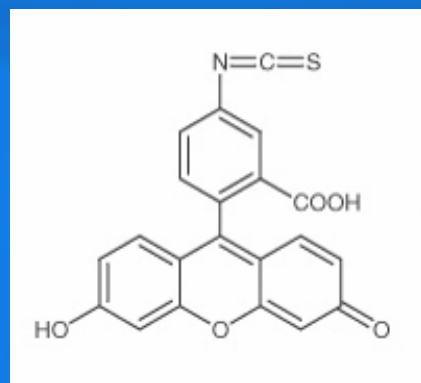
**Coumarins** - low extinction coefficients, phototoxic

**BOPIDYs** - non-polar and relatively insoluble in water

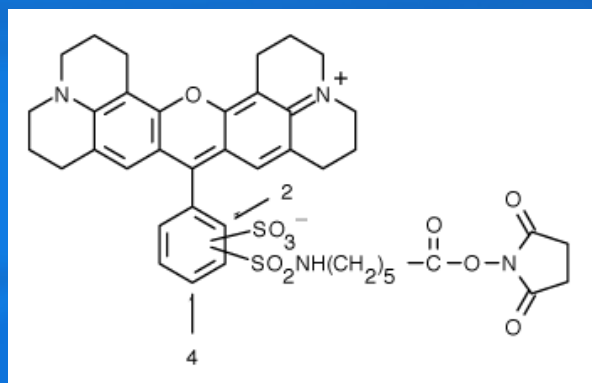
**Phthalocyanines** - photostable but difficult to synthesize and purify

**Cyanines** - high extinction coefficients and reasonable quantum yields

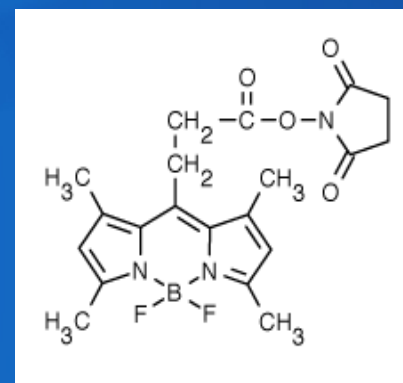
# Examples of Extrinsic Labels



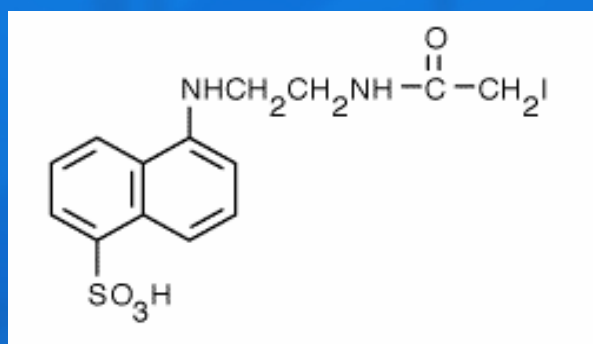
**FITC**  
(488/512),  $\tau \approx 4.0$



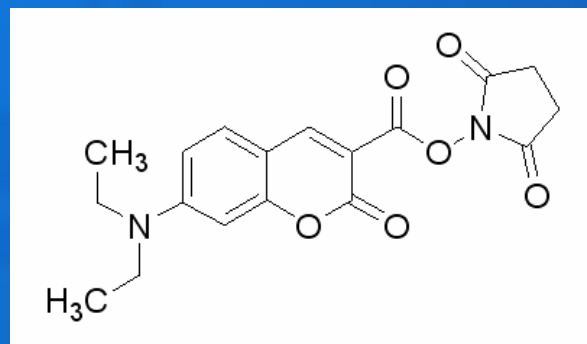
**Texas Red-NHS**  
(595-615),  $\tau \approx 3.5$  ns



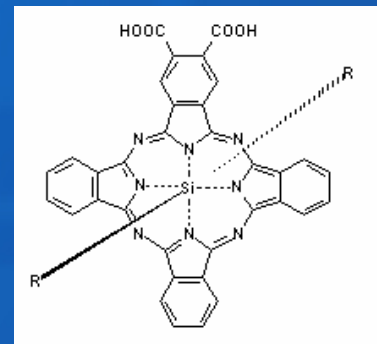
**BODIPY**  
(493/503),  $\tau = 6$  ns



**IAEDANS**  
(360/480),  $\tau \approx 15$  ns

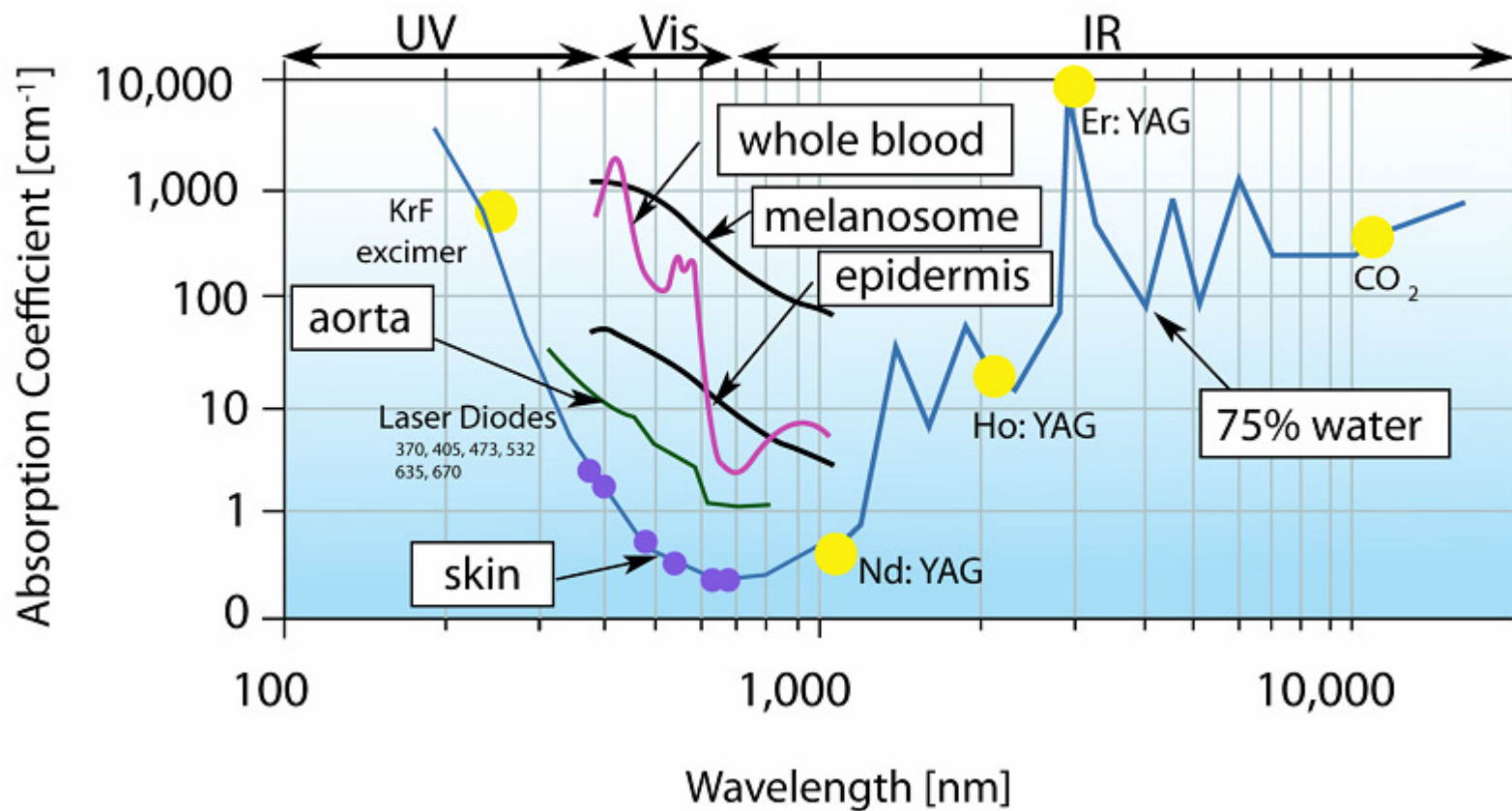


**Coumarin-3-carboxylic acid  
-NHS**  
(445/482),  $\tau \approx 2 - 3$  ns



**LaJolla Blue**  
(680/700)

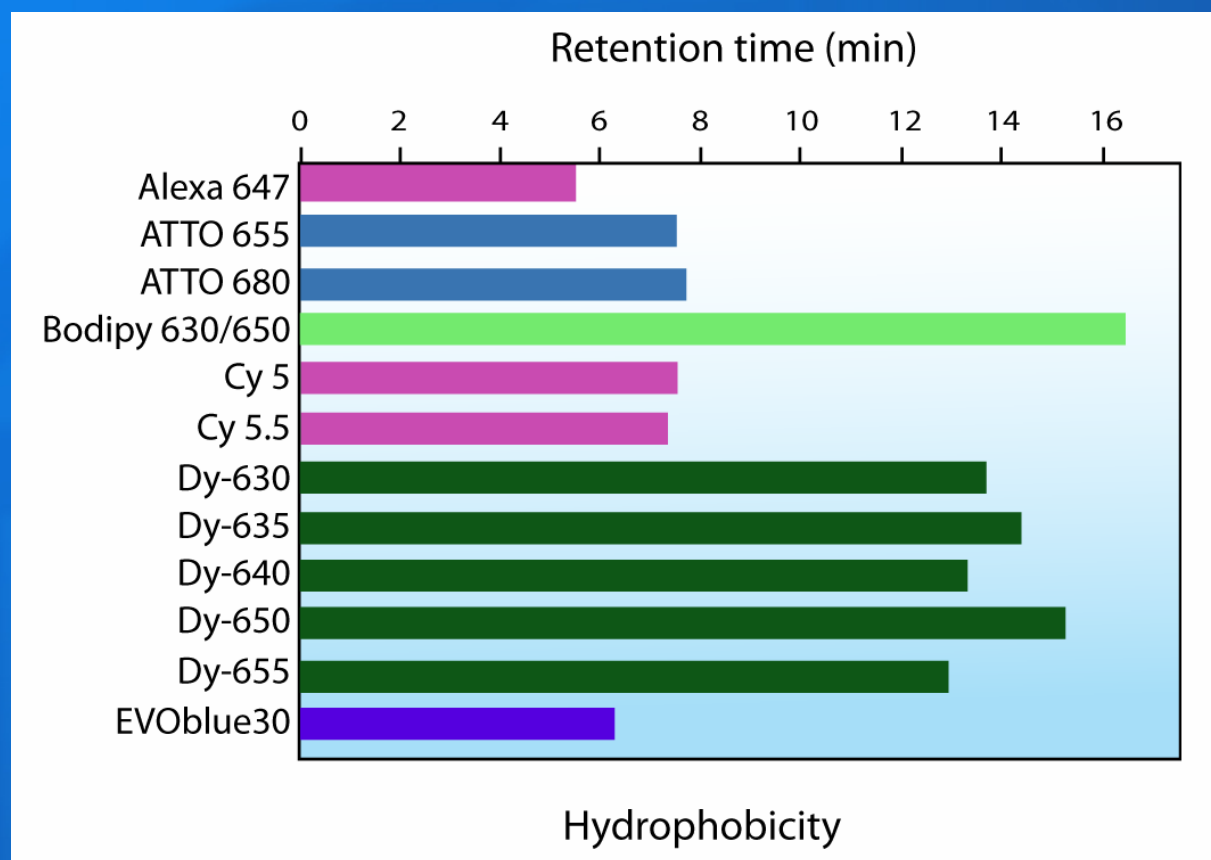
# Absorption of Biological Material



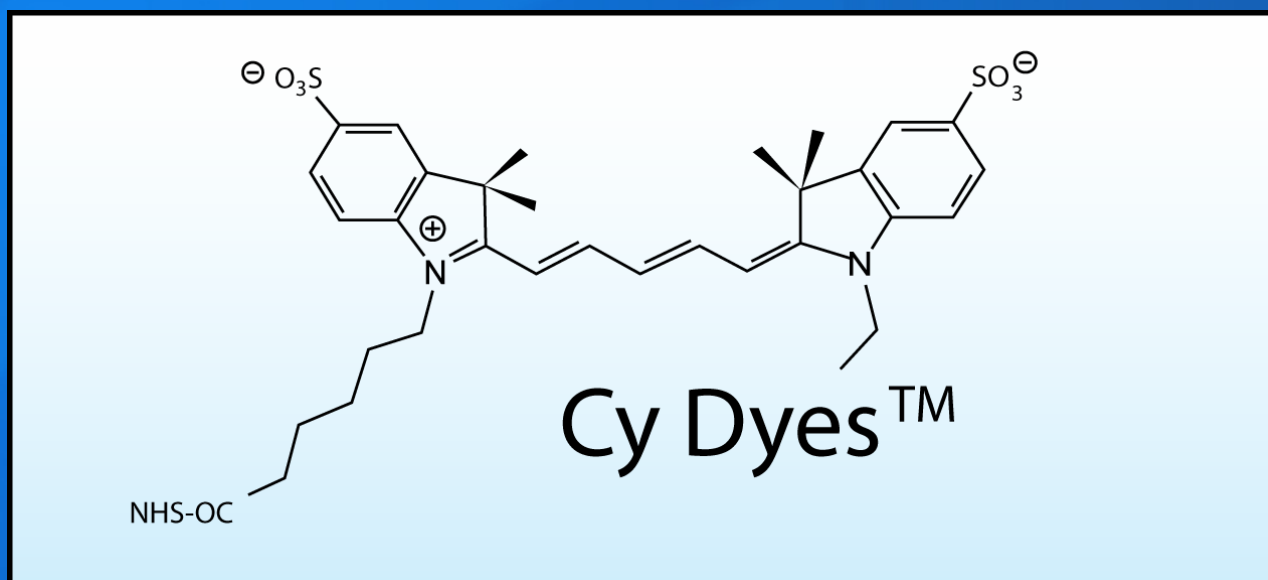
## Spectral Properties and Quantum Yields of Representative Long-Wavelength Absorbing and Emitting Dyes

Dye	$\lambda_{\max}$ (abs) [nm]	$\lambda_{\max}$ (em) [nm]	$\epsilon$ ( $M^{-1}cm^{-1}$ )	Lifetime $\tau$ [ns]	Q.Y. ( $H_2O$ )
Cy5	649	670	250,000	1.0	0.3
Dy650	649	670	120,000	0.64	0.05
Alexa 647	647	666	265,000	1.0	0.33
DyLight 649	646	674	250,000	1.0	0.33
HiLyte 647	649	674	250,000	1.0	0.28
BODIPY	629	646	97,000	3.9	0.7
Atto	661	678	100,000	1.8	0.3

# HPLC-Retention Times of Red Fluorescent Dyes

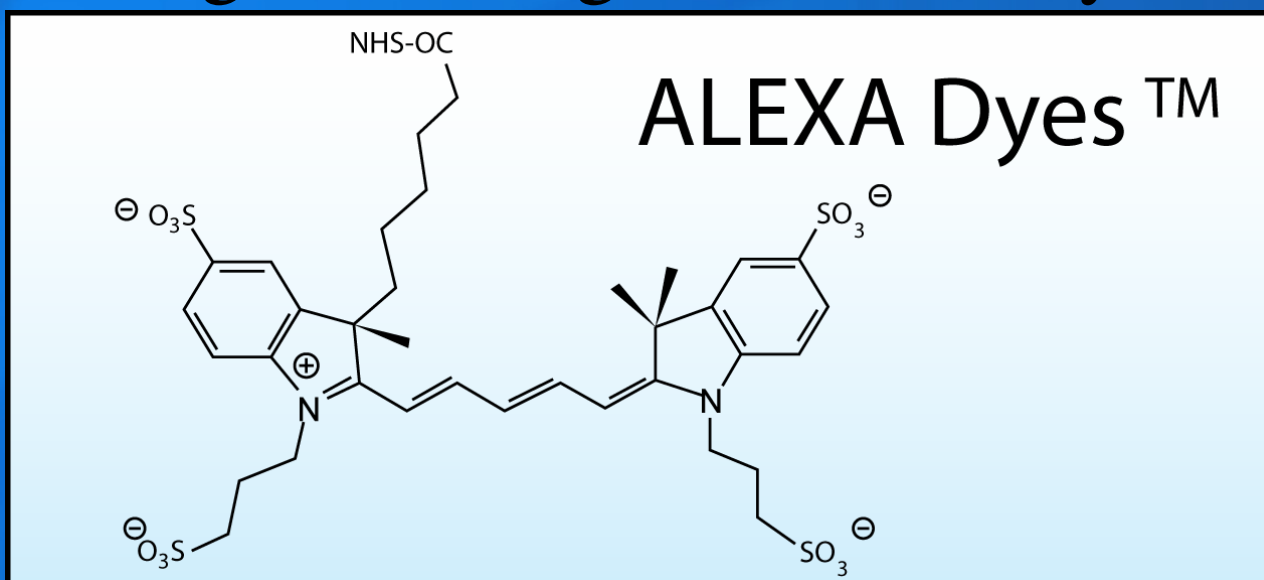


# Structure and Spectral Properties of Long-Wavelength Cy Dyes<sup>TM</sup>



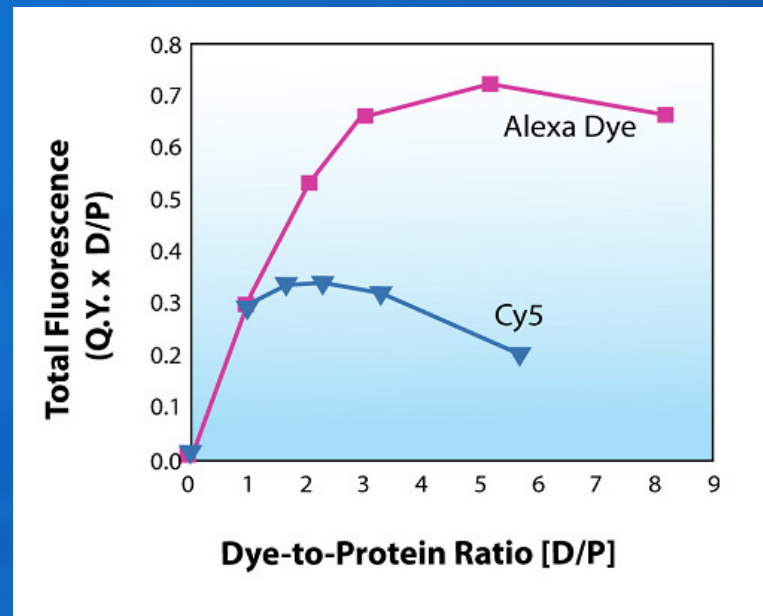
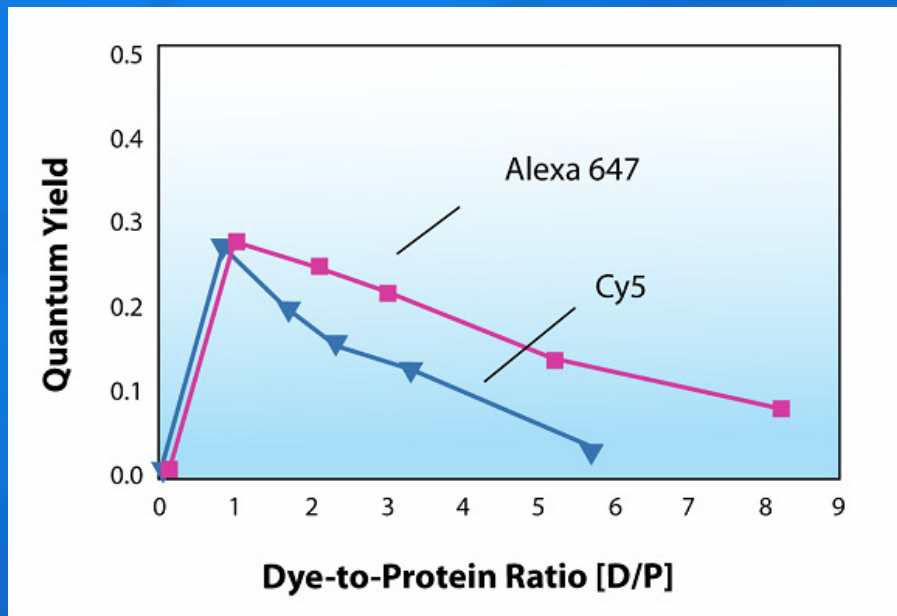
Dye-Conjugate	$\lambda_{\max}$ (abs) [nm]	$\lambda_{\max}$ (em) [nm]	$\epsilon$ ( $M^{-1}cm^{-1}$ )	Q.Y.
Cy5	649	670	250,000	0.2-0.28
Cy5.5	675	694	250,000	0.28
Cy7	743	767	250,000	0.28

# Structure and Spectral Properties of Long-Wavelength ALEXA Dyes™



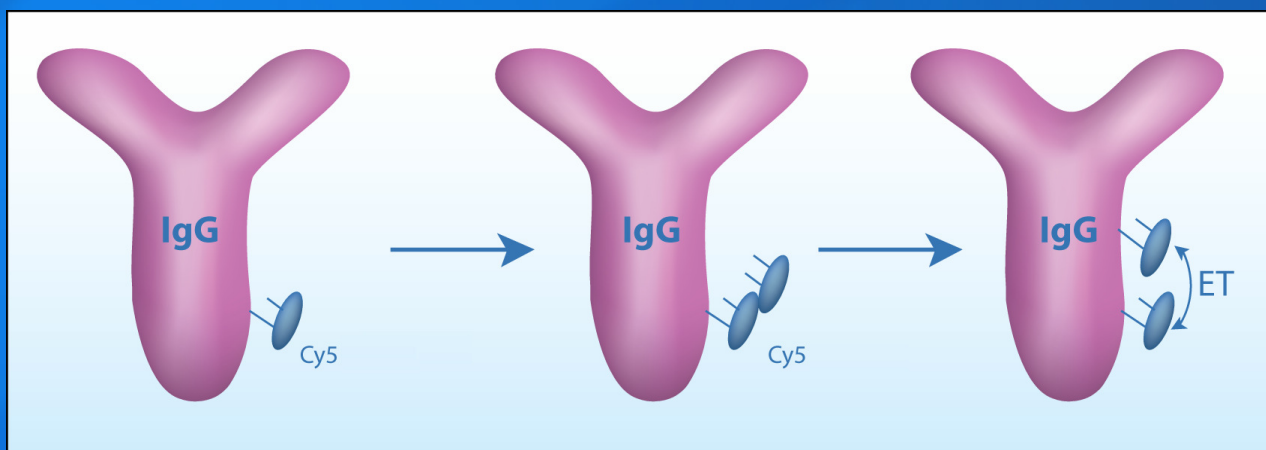
Dye-Conjugate	$\lambda_{\max}$ (abs) [nm]	$\lambda_{\max}$ (em) [nm]	$\epsilon$ ( $M^{-1}cm^{-1}$ )	Q.Y.
AF647	650	668	250,000	0.33
AF660	668	668	132,000	0.37
AF750	749	782	240,000	0.12

# Quantum Yields and Total Fluorescence of IgG-Conjugates of Cy5 (▼), and Alexa-647(■) for Various D/P Ratios



Cy5 has anomalous tendency to aggregate

# Cy5 - Anomalous Tendency to Aggregate

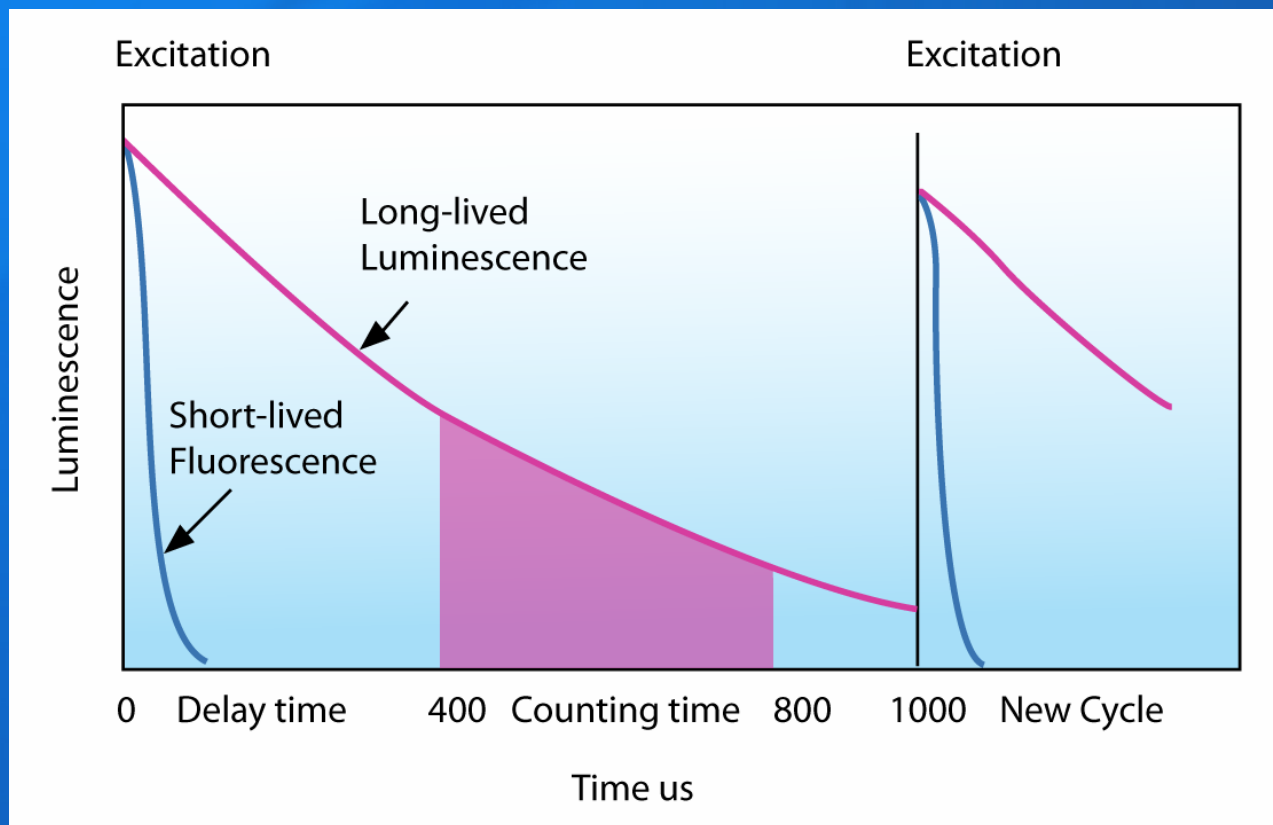


1<sup>st</sup> Cy5 molecule associates with the surface of a protein and reacts

2<sup>nd</sup> molecule interacts with labeled Cy5 and then reacts with next closest amino-group

Gruber et al., *Bioconjugate Chem.* **2000**, *11*, 696–704

# Time-Resolved Luminescence Measurement



# Luminescent Lanthanides ( $\text{Eu}^{3+}$ , $\text{Tb}^{3+}$ )

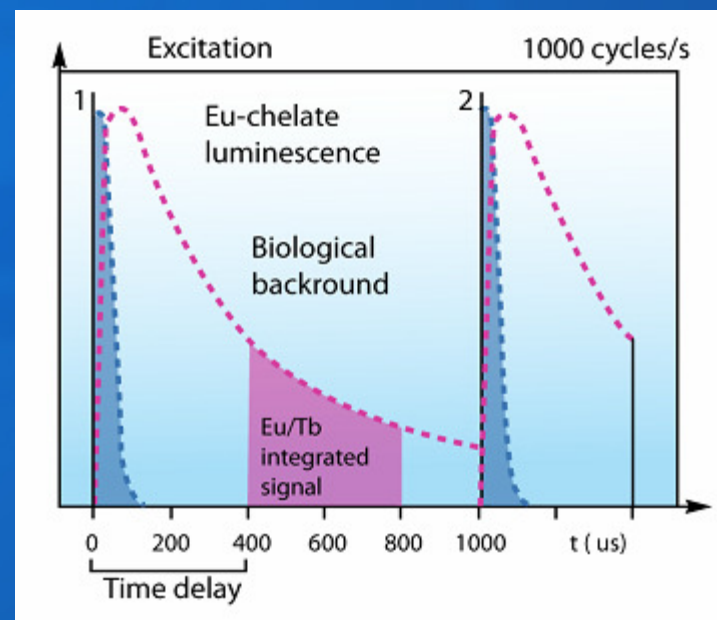
## Advantages vs. Organic Fluorophores:

Enable Discrimination of Short-Lived Autofluorescence by  
“Gating” - More Sensitive and Reliable Measurements

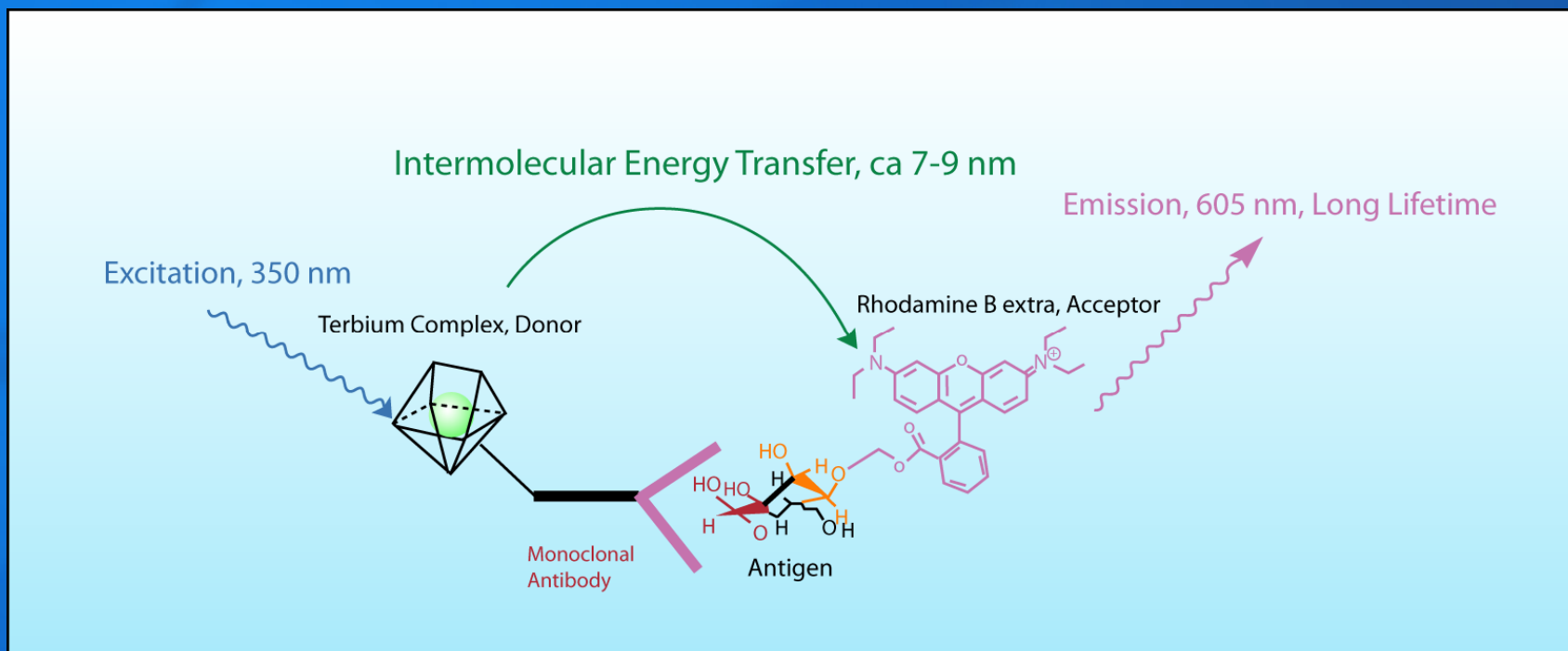
Narrow Emission Bands

Long Luminescent Lifetimes

Large Stokes' Shifts



# Homogeneous Time-Resolved Fluorescence Immunoassays (HTRF)



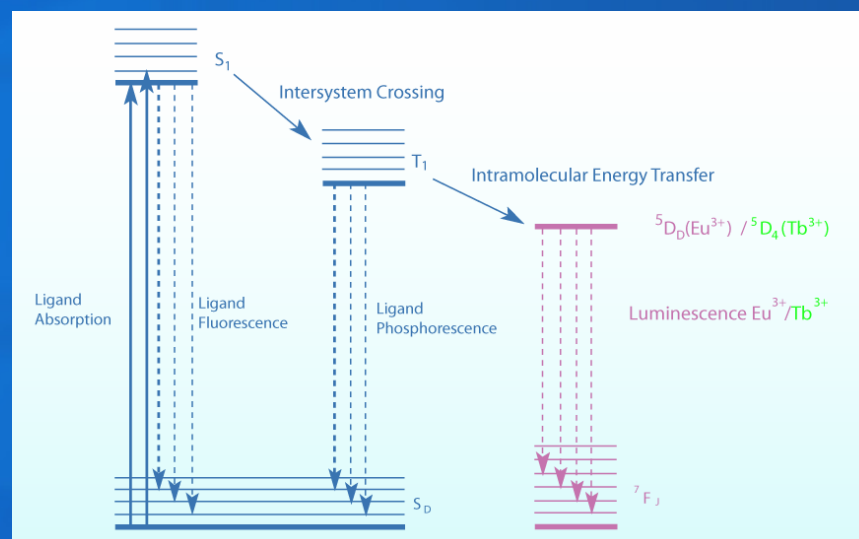
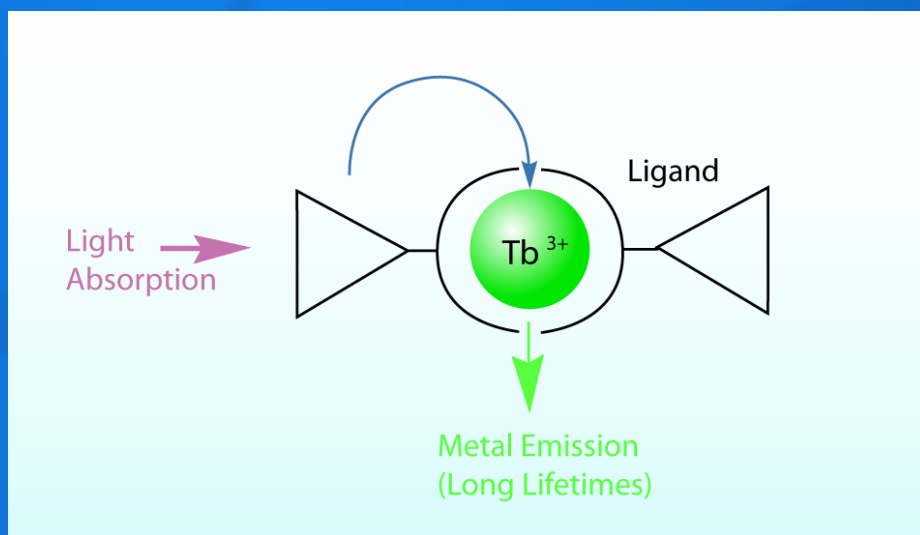
Minimal Sample Preparation - No Wash Steps

Applications: Drug Discovery

Clinical Diagnostics

# What Is The Mechanism?

## Antenna Effect

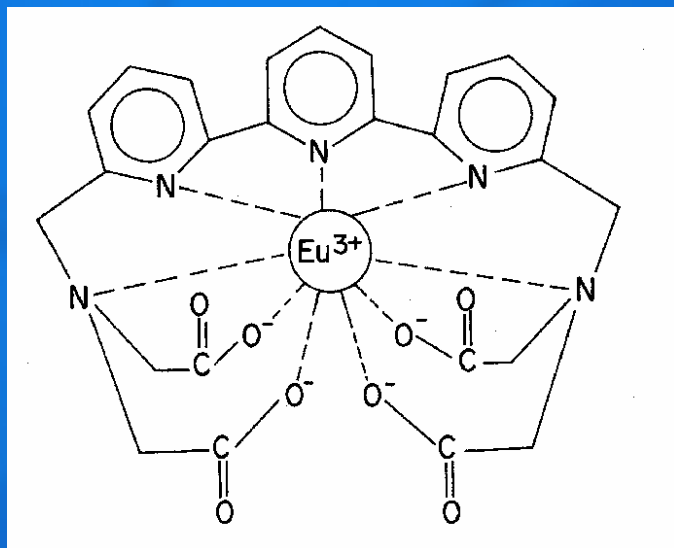


“Antenna Effect”: Strong Absorption and Good Energy Transfer

Ln(III) Protected from Quenching

Thermodynamically Stable and Kinetically Inert Complexes

# Lanthanide Complexes for Homogeneous Time-Resolved Immunoassays

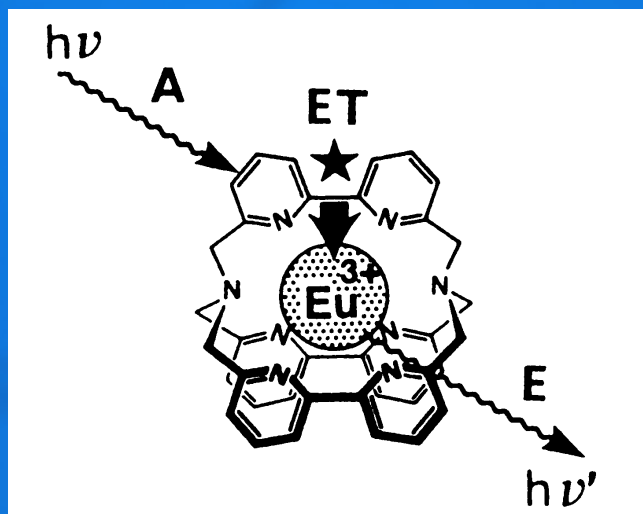


**LANCE Ultra™**



- Eu<sup>3+</sup> - Luminescence
  - $\phi = 11 - 15 \%$ ,  $\lambda_{\text{excitation}} \sim 320 \text{ nm}$
- 
- CH- and CH<sub>2</sub>-Group are Replaced by CD and CD<sub>2</sub>
  - Acceptors : Allophycocyanine (APC) or cyanine dyes

# Lanthanide Complexes for Homogeneous Time-Resolved Immunoassays



***Eu-Cryptate***<sup>TM</sup>

**Luminescent Only in Presence of  $\text{F}^-$**

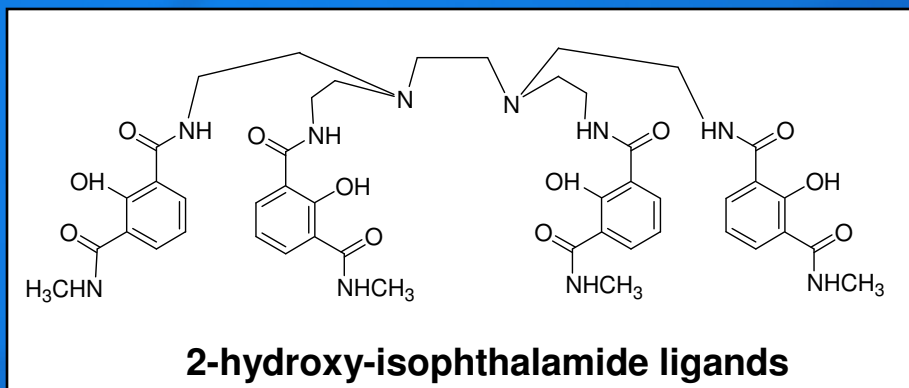
$$\phi = 2\%$$

Not Stable in Water but “Kinetically Inert”

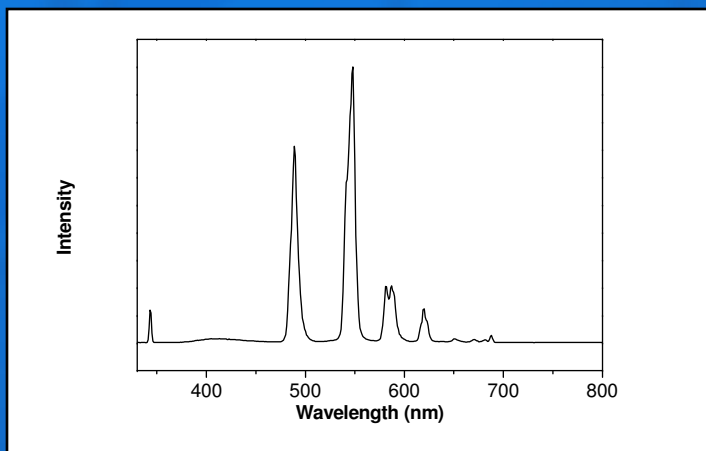
1-3 Water Molecules are Coordinated to Ln(III) (Limited Protection)

Acceptor : XL665 or C2 a low-MW acceptor

# Highly Luminescent Lanthanide Complexes



**Ligand**    **Tb<sup>3+</sup>**    **Eu<sup>3+</sup>**

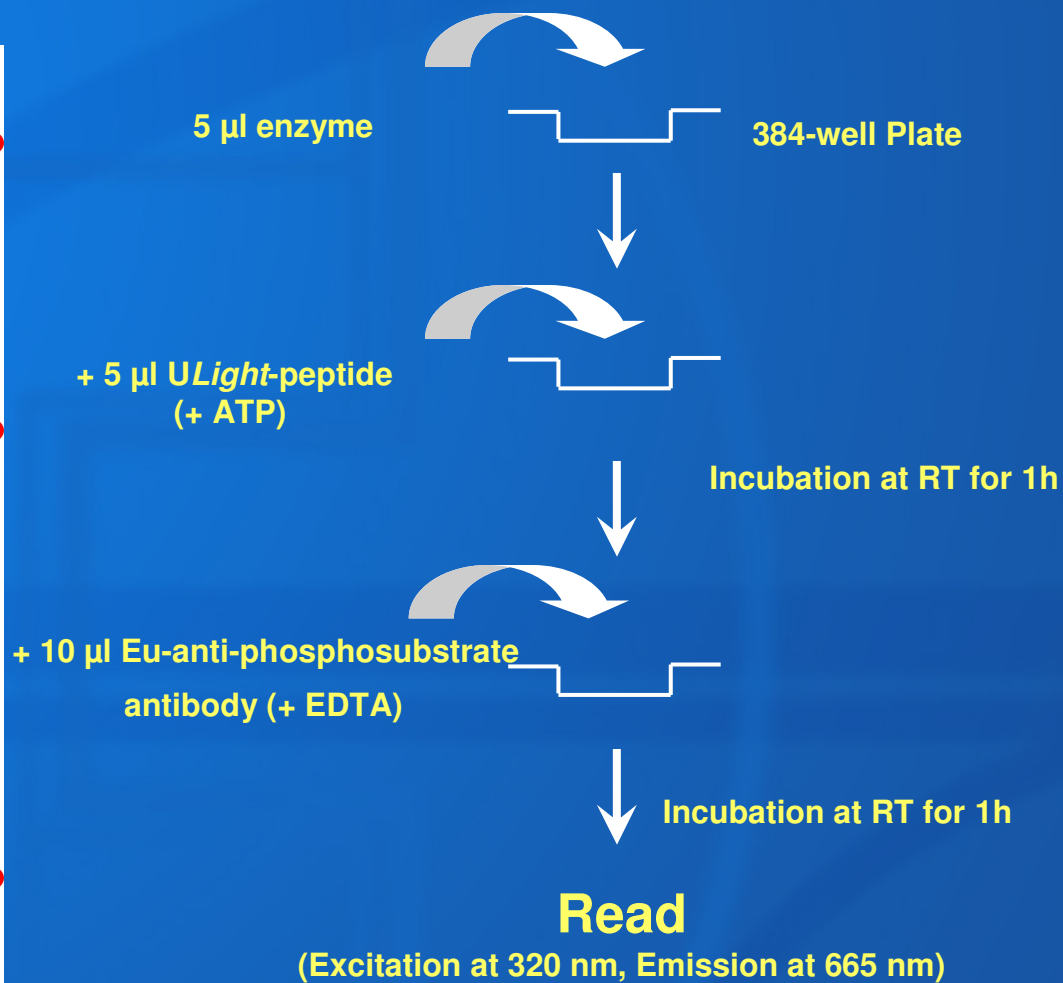
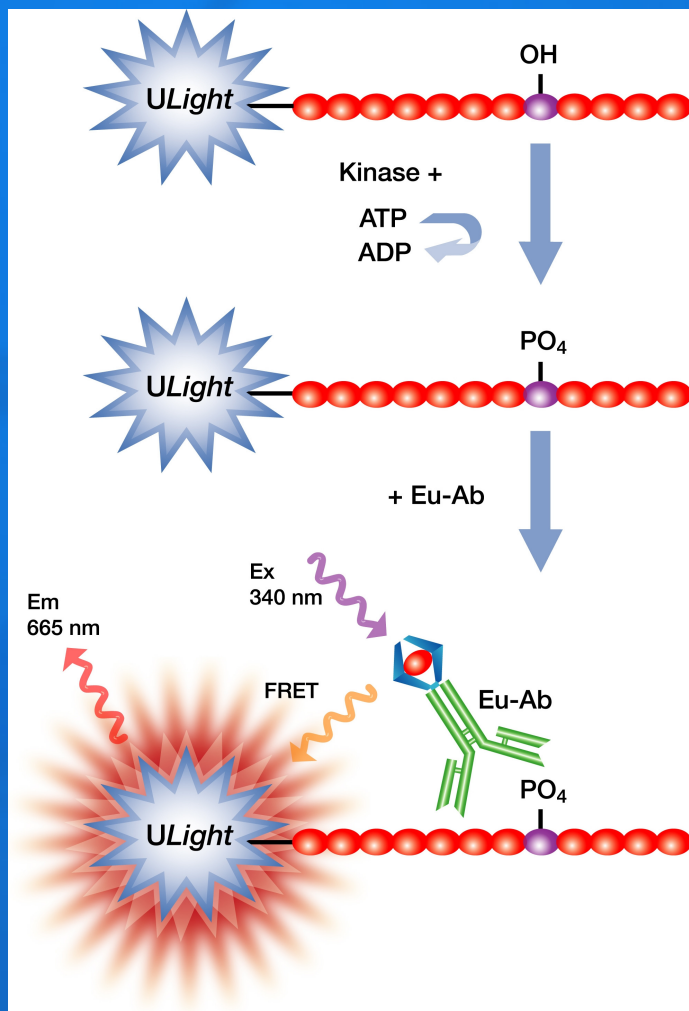


**Lumiphore, Inc.**

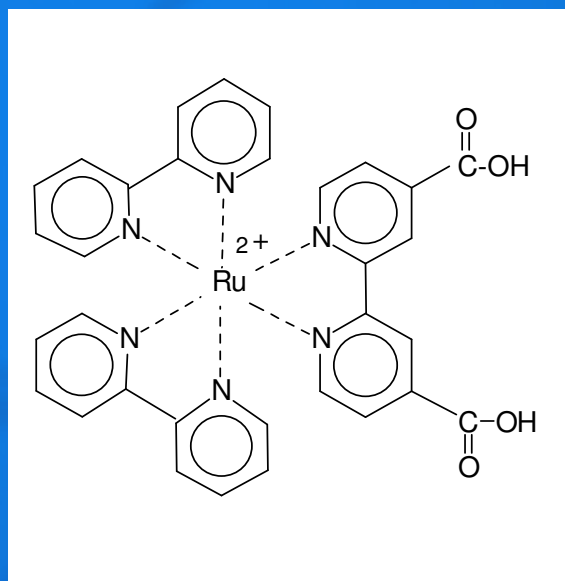
**Quantum Yield: 60 % !!!**

**Lifetime  $\tau = 2.56$  ms**

# TR-FRET HTS Assay



# Spectral Data and Lifetimes for a Representative Metal-Ligand Complexes (MLCs)



**Ru(bpy)<sub>2</sub>(dcbpy)**

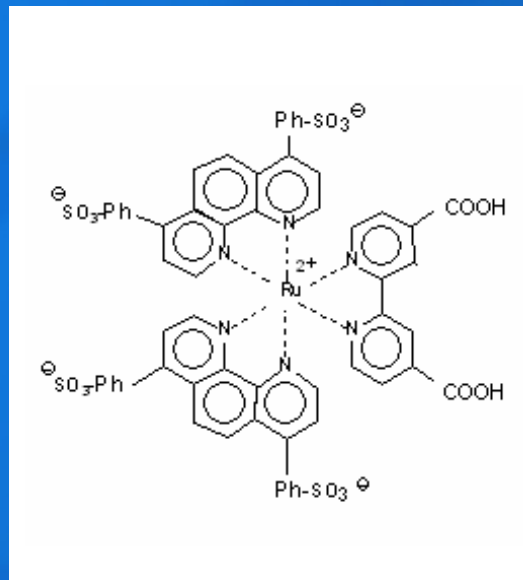
$\lambda_{\max}(\text{abs})$  (water) = 467 nm

$\lambda_{\max}(\text{em})$  = 655 nm

Q.Y. = 0.05

$r_0$  = 0.23

$\tau$  = 366 ns



**Ru(SO<sub>3</sub>dphphen)<sub>2</sub>(dcbpy)**

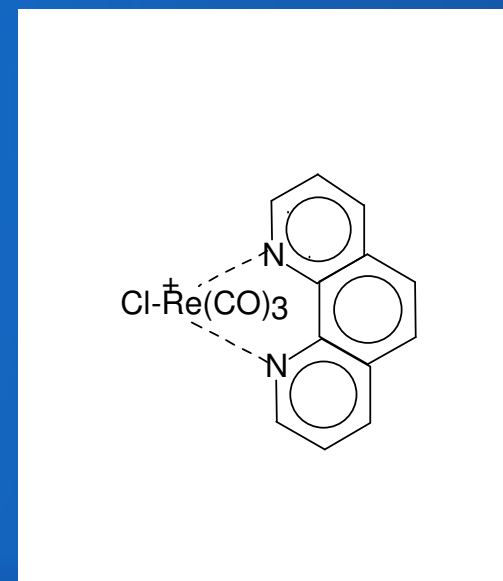
$\lambda_{\max}(\text{abs})$  (water) = 4640 nm

$\lambda_{\max}(\text{em})$  = 643 nm

Q.Y. = 0.06

$\tau$  = 0.8  $\mu\text{s}$

$\tau$  = 2  $\mu\text{s}$  (HSA)



**Re(CO)<sub>3</sub>Cl(phen)**

$\lambda_{\max}(\text{abs})$  (water) = 275 nm

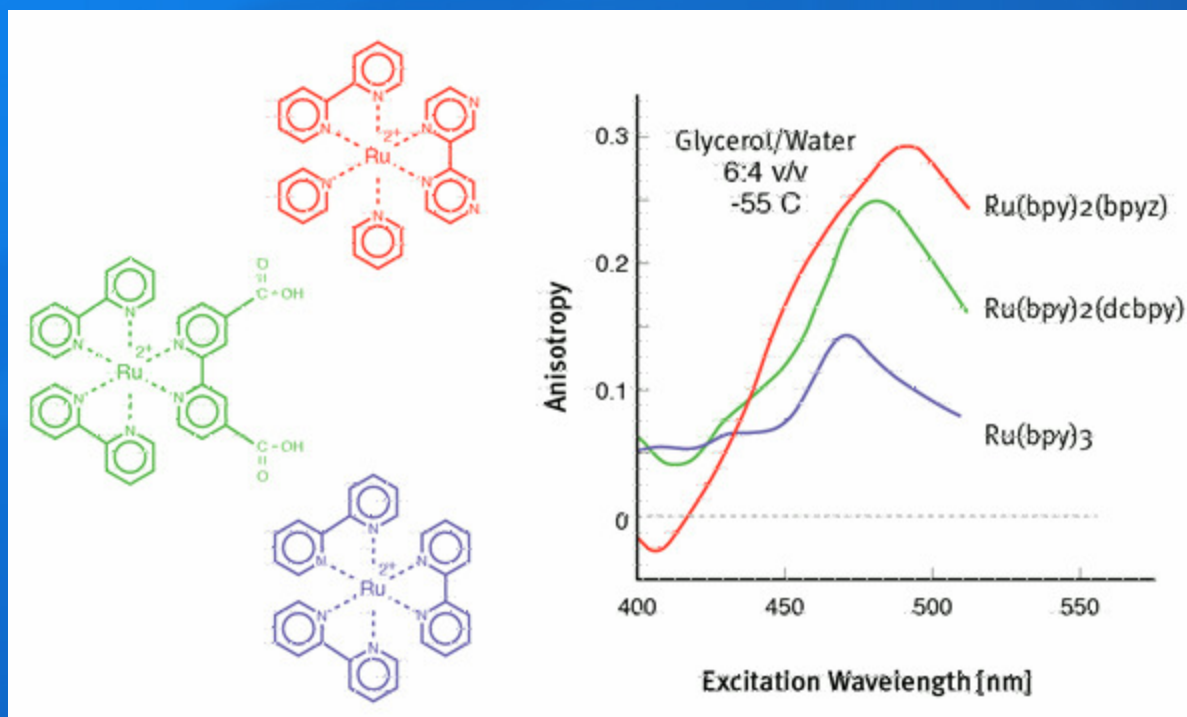
$\lambda_{\max}(\text{em})$  = 589 nm

Q.Y. = 0.2

$r_0$  = 0.3

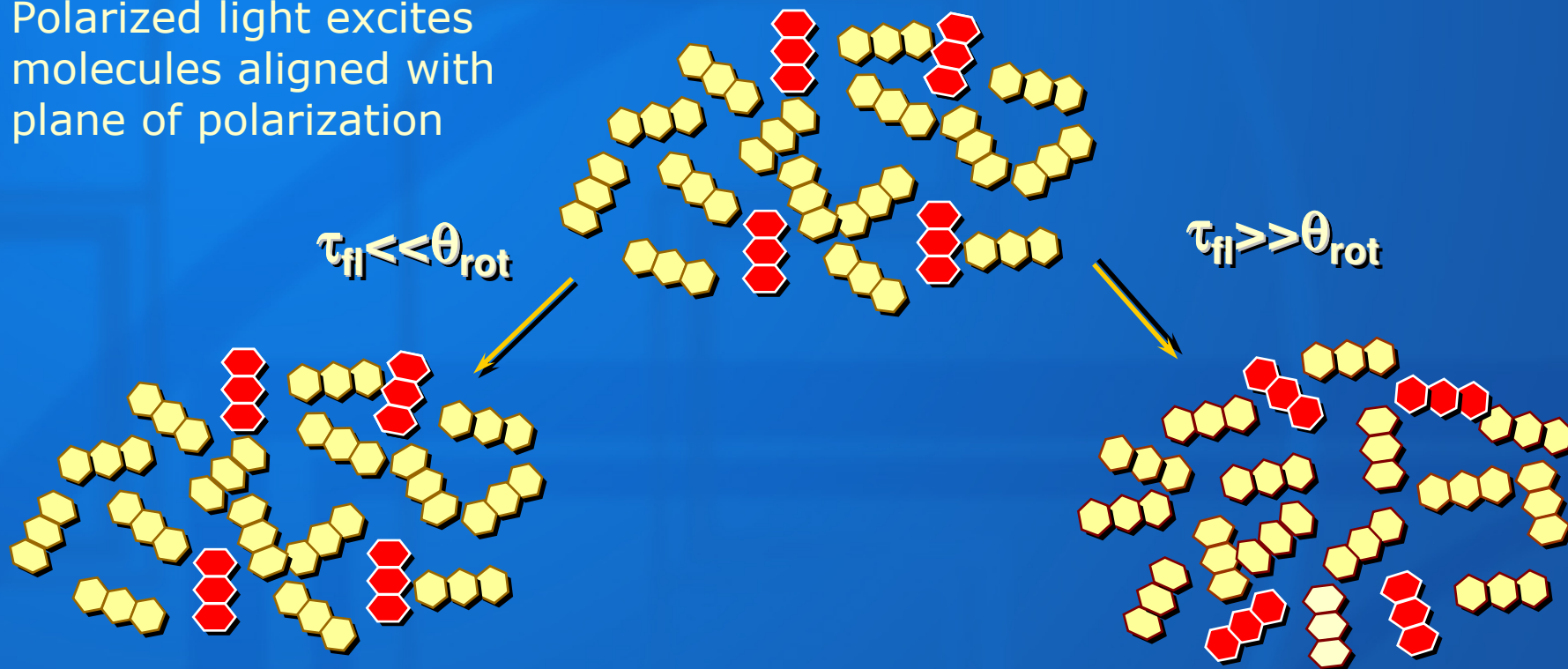
$\tau$  = 110 ns

# Excitation Polarization Spectra of Representative MLCs



# Fluorescence Polarization: A Race between Emission and Molecular Motion

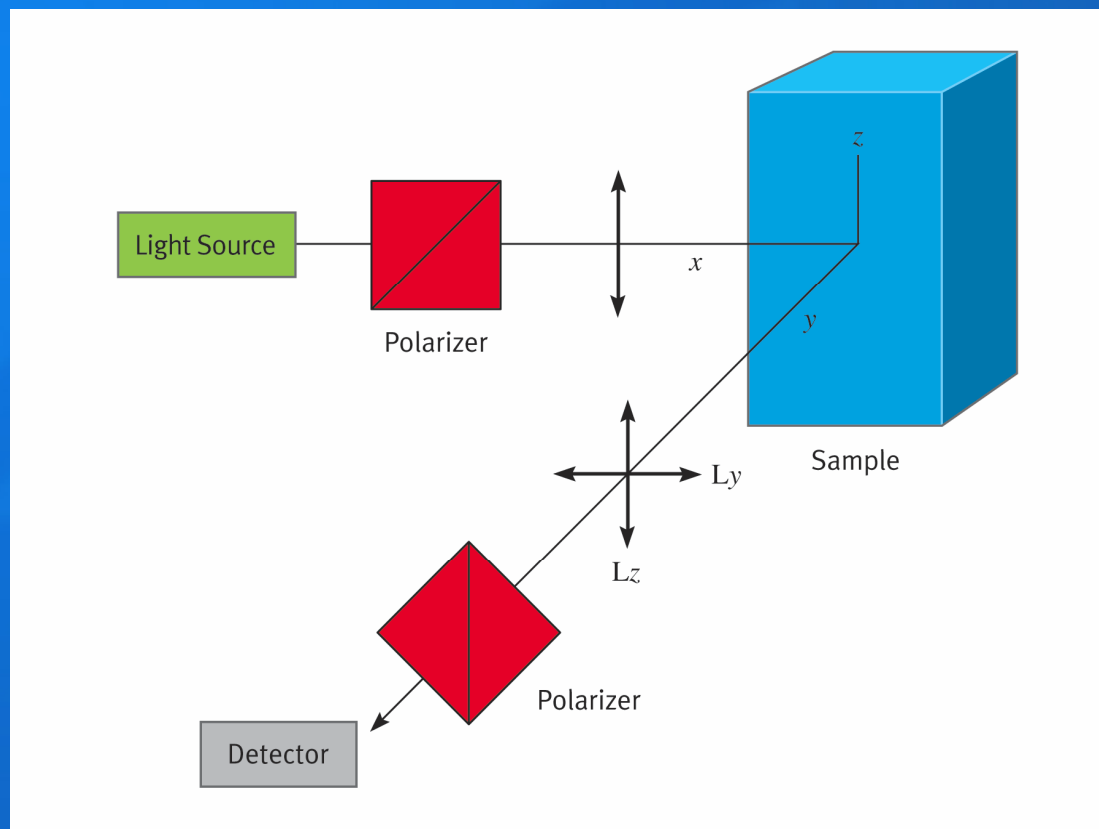
Polarized light excites  
molecules aligned with  
plane of polarization



Excited molecules remain aligned.  
Fluorescence is polarized.

Orientation of excited molecules  
randomizes. Fluorescence is depolarized.

# Fluorescence Polarization Measurement



## Fluorescence Polarization

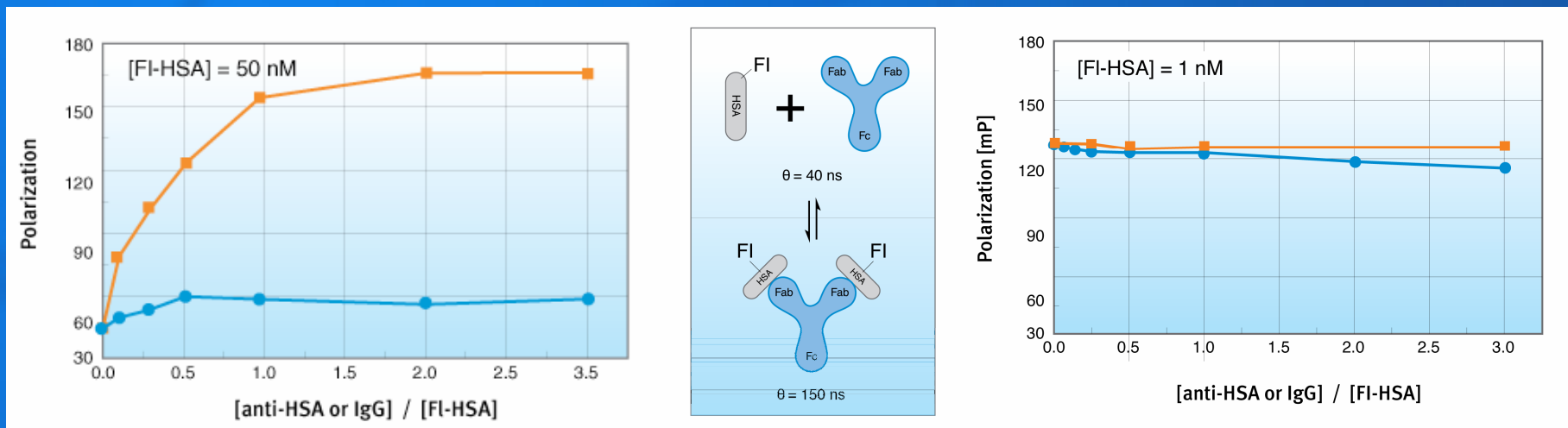
$$\text{Polarization (P)} = \frac{I_v - I_h}{I_v + I_h}$$

$$\text{Anisotropy (r)} = \frac{I_v - I_h}{I_v + 2 I_h}$$

$$P = \frac{3 r}{2 + r}$$

$$r = \frac{2 P}{3 - P}$$

# Role of Lifetime in FP



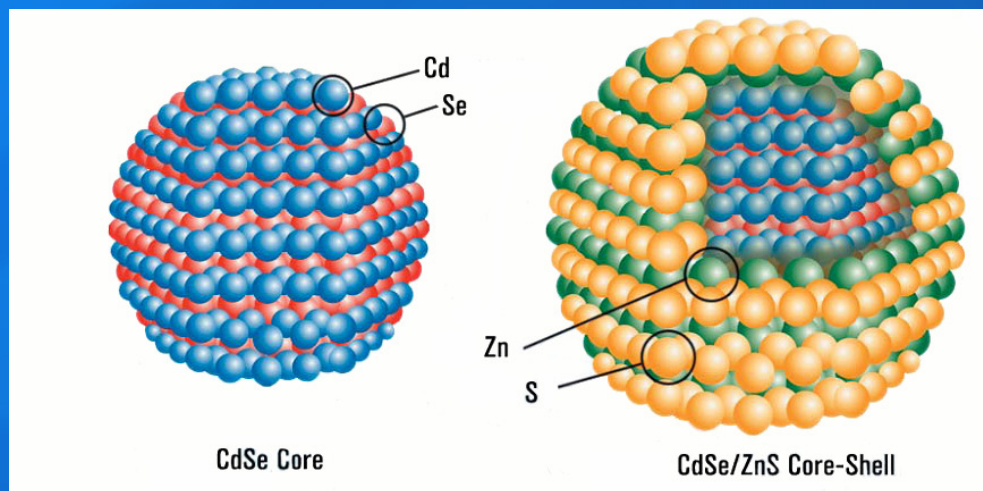
$$r = \frac{r_0}{(1 + \tau/\theta)}$$

$$\tau_{fl} \sim 300 \text{ ns}$$

$$\tau_{fl} = 4 \text{ ns}$$

$$\theta = 150 \text{ ns}$$

# Quantum Dots



# Quantum Dots

## Nanometer-Scale Atom Clusters

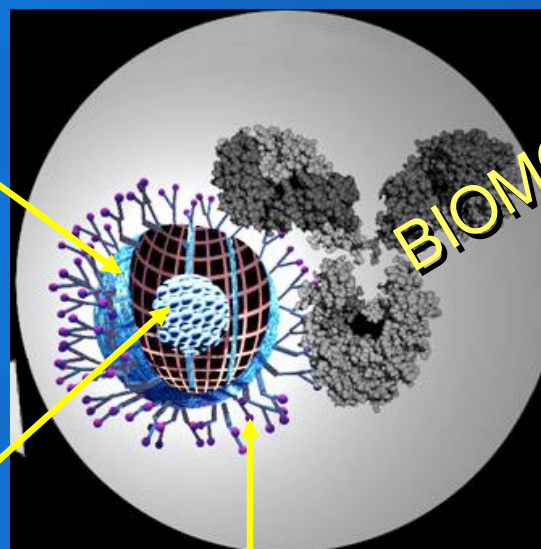
**CORE**

Cadmium selenide (**CdSe**), or  
Cadmium telluride (**CdTe**)  
few hundred – few thousand atoms

The semiconductor material is chosen  
based upon the emission wavelength,  
however it is the **size** of the particles  
that **tunes the emission  
wavelength**.

**SHELL**

In the core emission is typically weak  
and always unstable.  
The shell material (**ZnS**) has been  
selected to be almost entirely  
unreactive and completely insulating  
for the core.

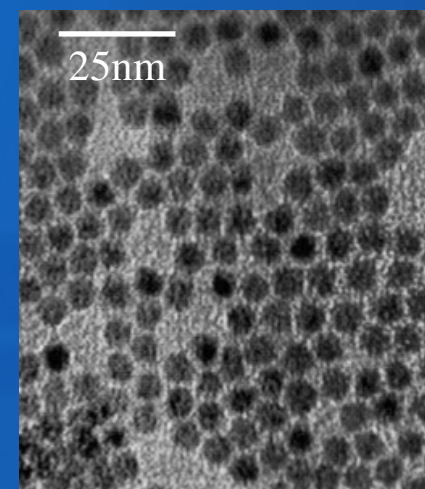
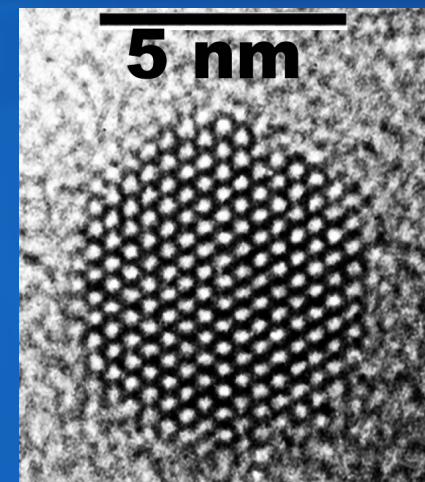
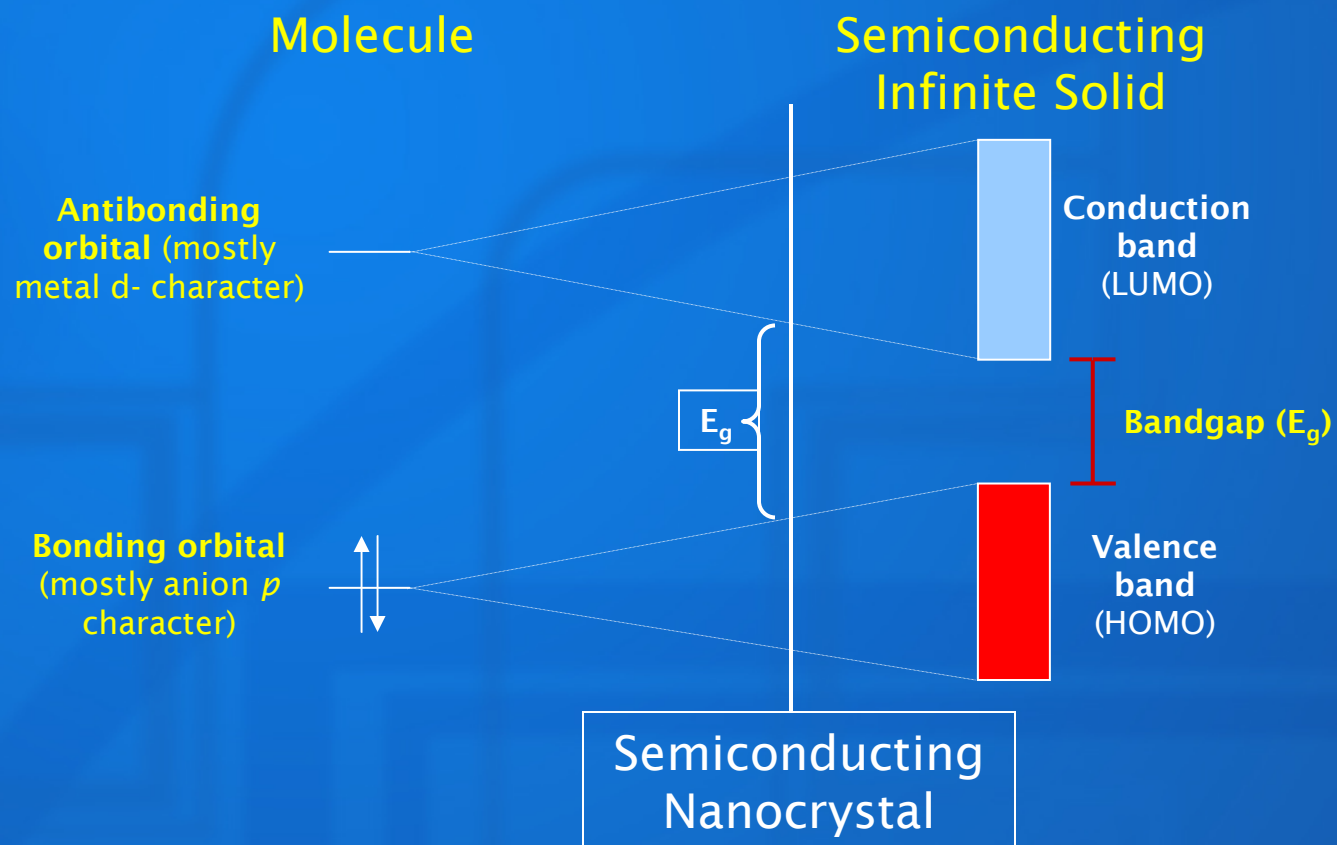


**BIOMOLECULE**

**COATING**

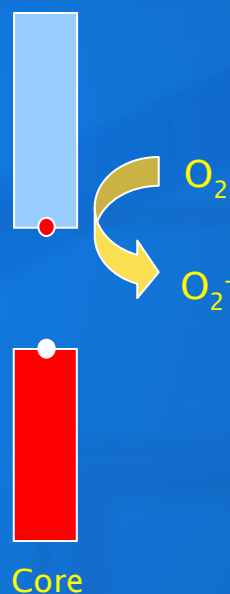
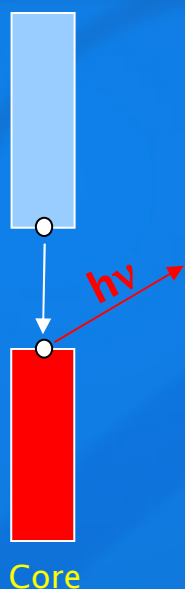
A layer of organic ligands covalently attached to the surface  
of the shell. This coating provides a **surface for  
conjugation** to biological (antibodies, streptavidin, lectins,  
nucleic acids) and nonbiological species and makes them  
“water-soluble”

# Semiconductor Nanocrystals

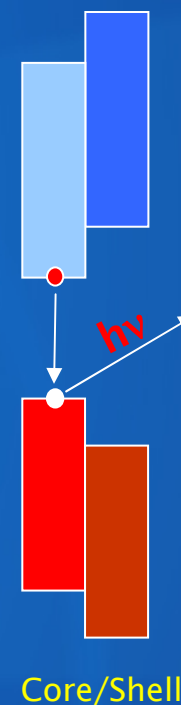


Bandgap of nanocrystal is size-dependent, larger than for bulk material

# Preventing Photobleaching In Quantum Dots



Plain core QDots show emission, but oxidation results in permanent loss of emission, so the population would gradually bleach away.

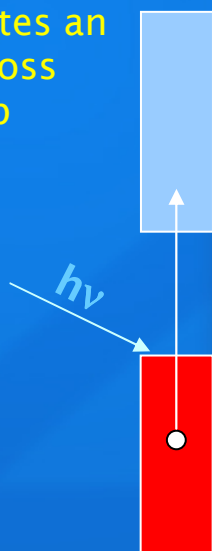


In Core/shell QDots the electron remains in the lower-energy core orbitals, and never reaches particle surface to react

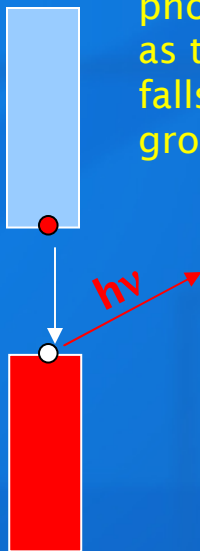
**The shell keeps the high-energy excited electron away from oxygen**

# Semiconductor Nanocrystal Fluorescence

A high-energy photon excites an electron across the bandgap



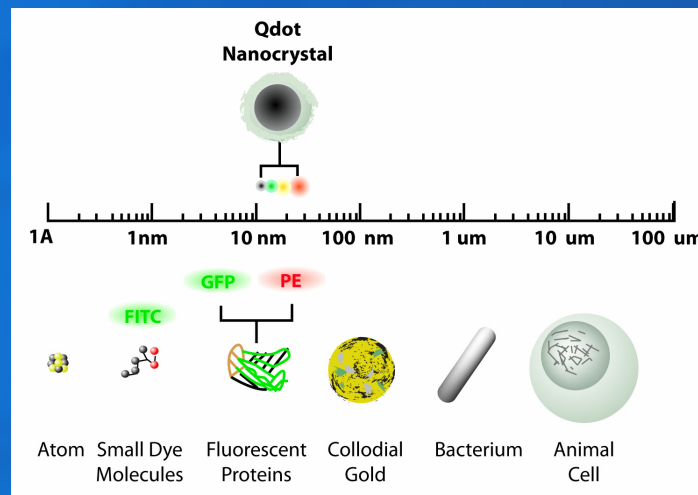
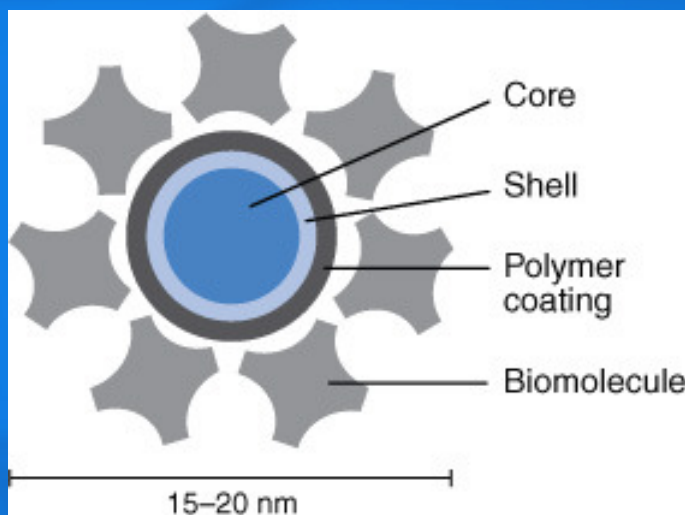
A bandgap-energy photon is emitted as the electron falls back to the ground state



Size-dependent bandgap means size-dependent color

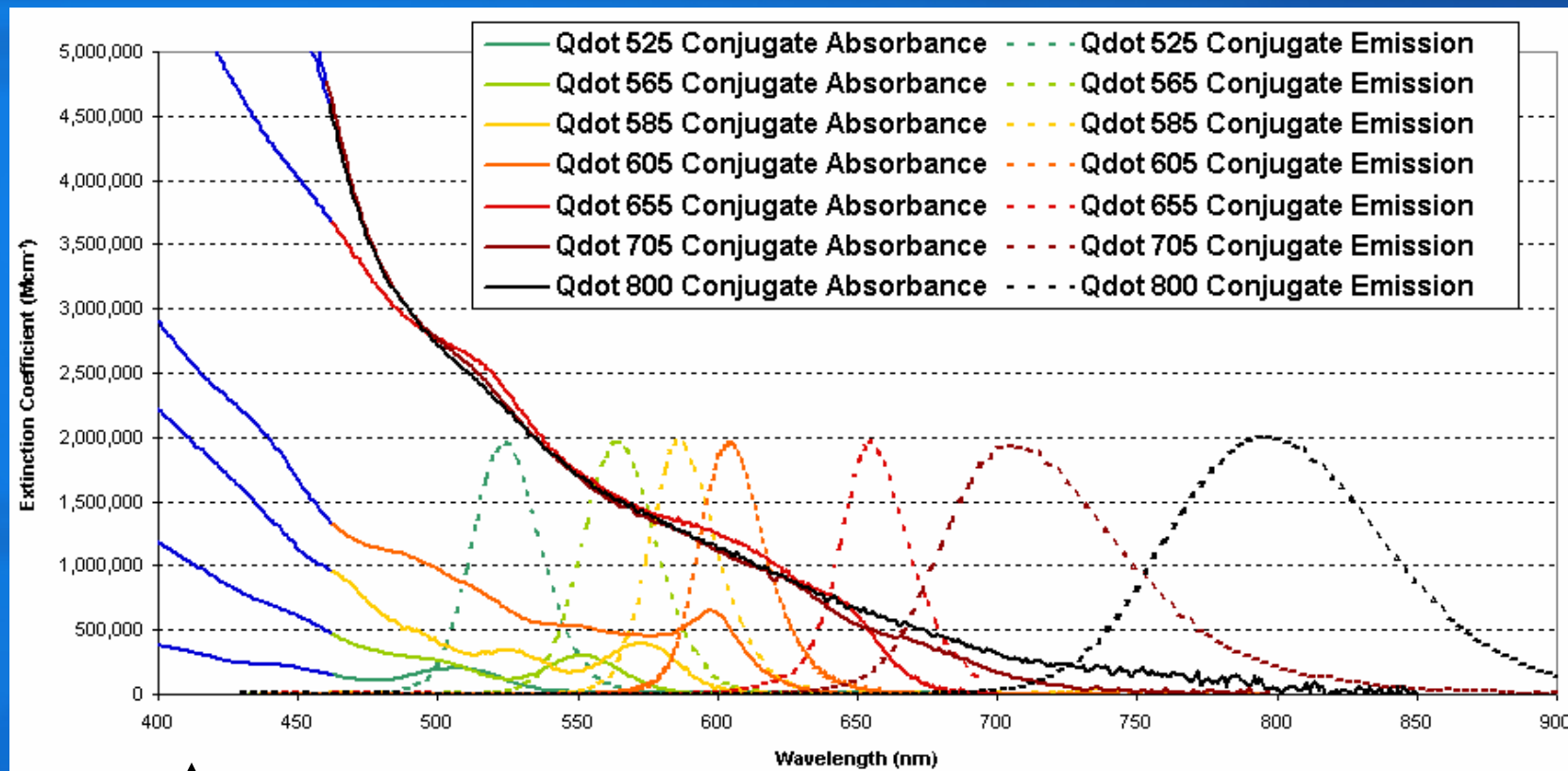
# Quantum Dots

## Nanometer-Scale Atom Clusters



Quantum Dot Material System	Emission Range	Quantum Dot Diameter Range	Quantum Dot Type	Standard Solvents	Example Applications
CdSe	465nm - 640nm	1.9nm - 6.7nm	Core	Toluene	Research, Solar Cells, LEDs
CdSe/ZnS	490nm - 620nm	2.9nm - 6.1nm	Core-Shell	Toluene	VisibleFluorescence Applications, Electroluminescence, LEDs
CdTe/CdS	620nm - 680nm	3.7nm - 4.8nm	Core-Shell	Toluene	Deep Red Fluorescence Apps.

# Qdot Optical Spectra



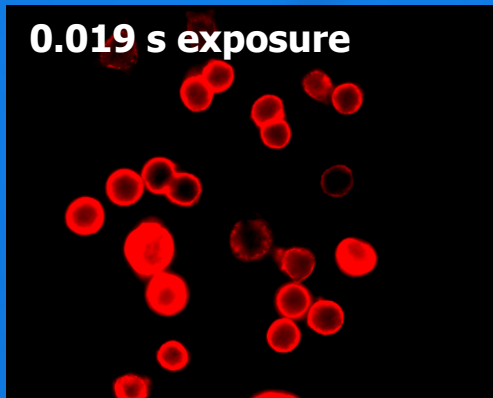
Violet  
excitation

Absorbance × Quantum Yield = **Brightness**  
 photons in      fraction converted      photons out

Broad range of emissions

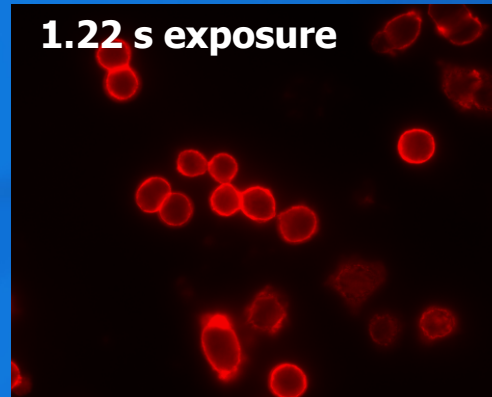
High absorbance means increased brightness  
 Single-color excitation, multicolor emission for easy multiplexing

## Qdot<sup>®</sup> nanocrystals



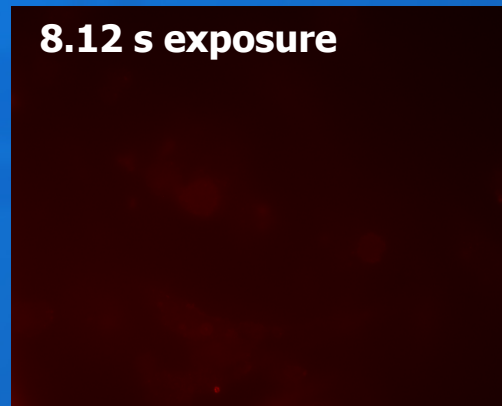
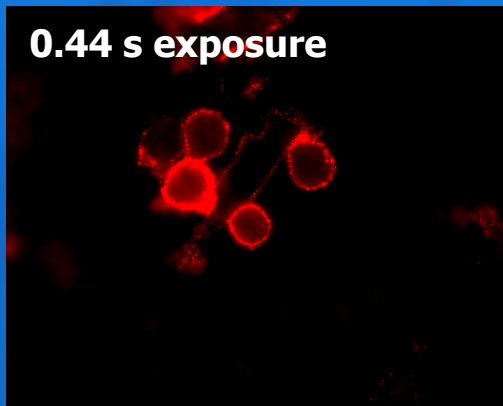
60X  
↔

## Cy5 organic dye



## Anti-Her2/neu + anti-mouse Ig conjugates

- SK-BR-3 Cells: High Her2/neu expression
- Nanocrystals up to 50x brighter



- MDA-MB-231 cells: Low Her2/neu expression
- Nanocrystals easy to detect but dye undetectable

# Photostability in Microscopy

## 3T3 Cells

### Top panel (a-e)

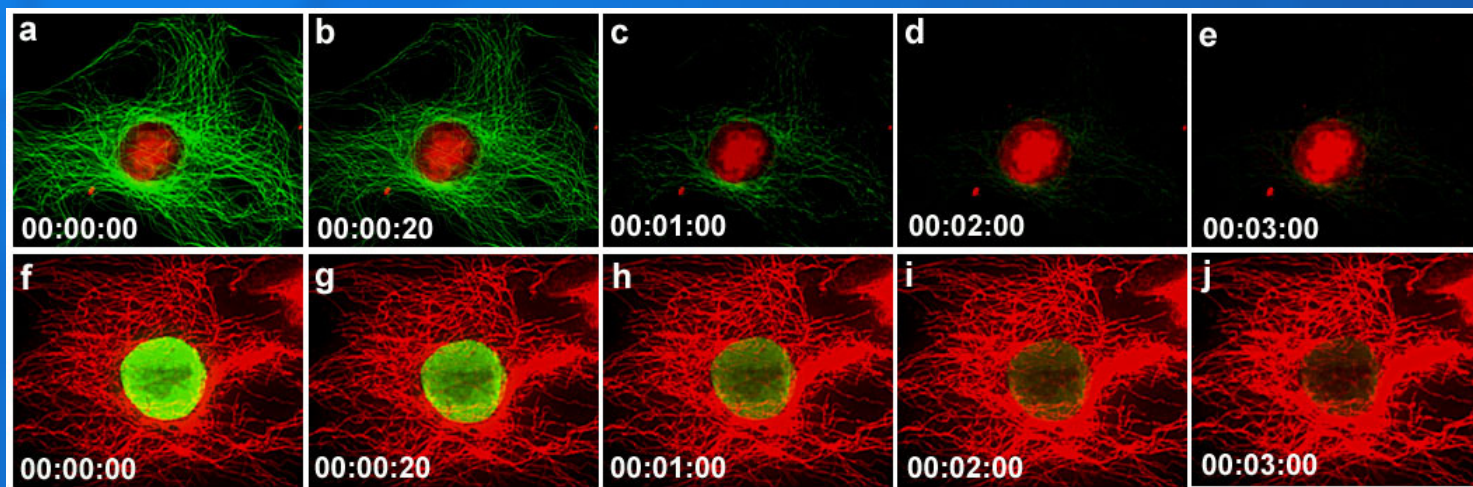
Nucleus: Qdot<sup>®</sup> 605 conjugate

Microtubules: Alexa Fluor<sup>®</sup> 488 conjugate

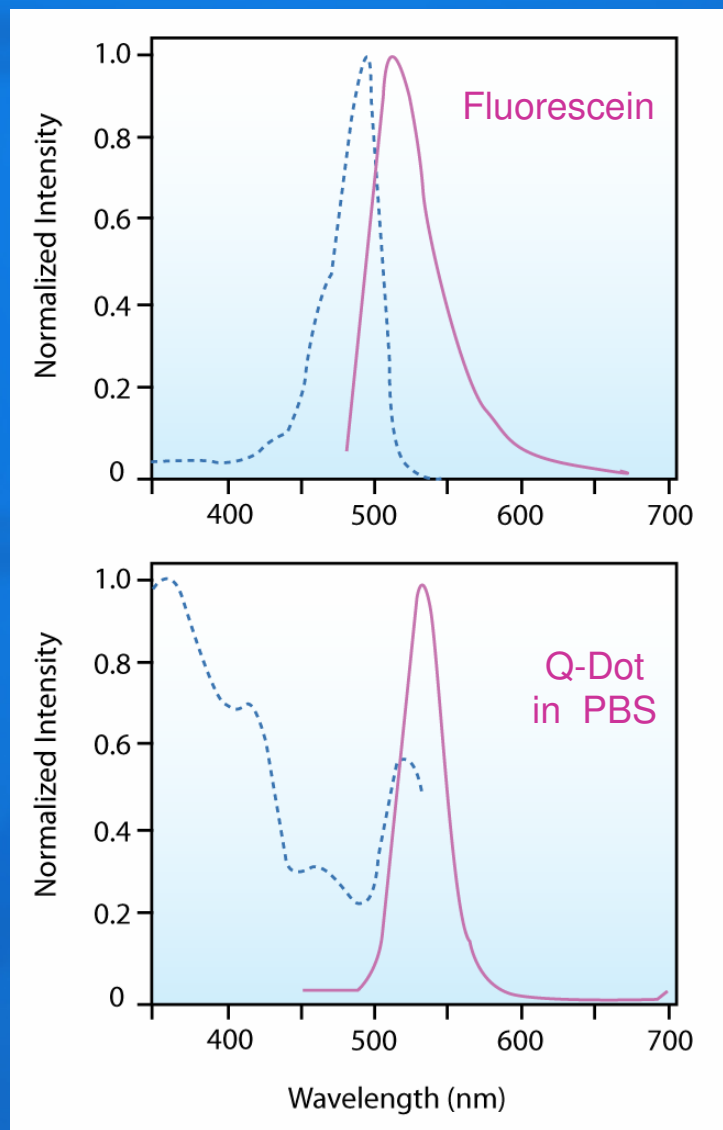
### Bottom panel (f-j)

Nucleus: Alexa Fluor<sup>®</sup> 488 conjugate

Microtubules: Qdot<sup>®</sup> 605 conjugate



**Photostability results in sensitivity, ease of use, and sample permanence**



## Advantages:

Broad absorption spectra, making it possible to excite all colors of QDs simultaneously with a single light source - **Multiplexing**

Narrow and symmetrical emission spectra

**Emission tunable** with size and material composition

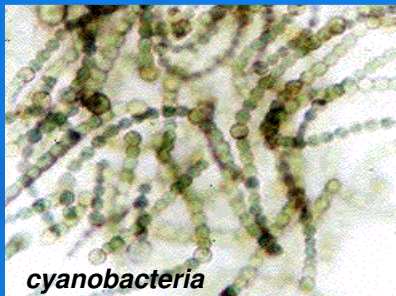
Exhibit excellent **photo-stability**

## Disadvantages:

**Large size and high mass** limit their use in applications requiring high diffusional mobility

QDot	$\lambda_{\max}(\text{abs})$ [nm]	$\lambda_{\max}(\text{em})$ [nm]	$\epsilon$ ( $\text{M}^{-1}\text{cm}^{-1}$ )	Q.Y.
655	350	655	9,000,000	~0.5
705	350	705	13,000,000	~0.5
800	350	800	13,000,000	~0.5

## Phycobiliproteins



From red algae and cyanobacteria (blue-green algae).

Absorb strongly between 470 and 650 nm.

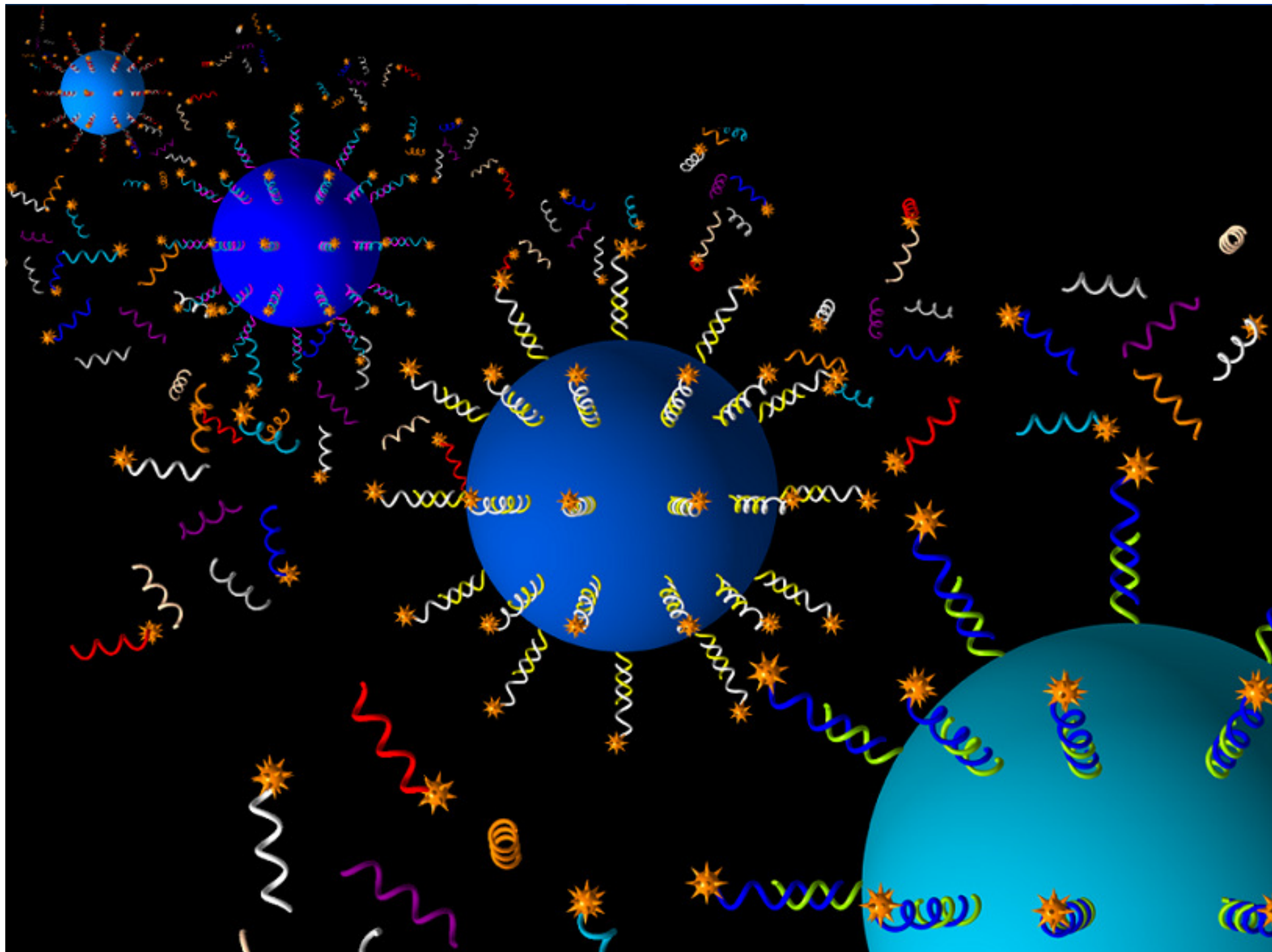
Highly fluorescent *in vitro*

Hold the record for the highest extinction coefficients and largest number of photons emitted before bleaching

Used as ET acceptors and in tandem-constructs for multi-color analysis (flow cytometry)

Four main classes of phycobiliproteins

Protein	Subunit Composition	Approx mol. wt.	$\epsilon (M^{-1}cm^{-1})$	Total bilins per protein	$\lambda_{max} (abs) [nm]$	$\lambda_{max} (em) [nm]$
Allophycocyanin	$(\alpha\beta)_3$	100,000	700,000	6	650	660
R-Phycocyanin	$(\alpha\beta)_3$	110,000	1,000,000	9	555,618	642
B-Phycoerythrin	$(\alpha\beta)_{6V}$	240,000	2,400,000	34	543,562	576
R-Phycoerythrin	$(\alpha\beta)_{6V}$	240,000	2,200,000	34	495,536,565	576



The background features a vibrant rainbow gradient from red on the left to green on the right. Scattered throughout are several blue spheres of varying sizes, each with numerous colorful, wavy lines radiating from them, resembling a suspension array. The word "Luminex" is written in a bold, white, italicized font in the upper right corner, with a small red dot above the letter 'i'.

***Luminex***

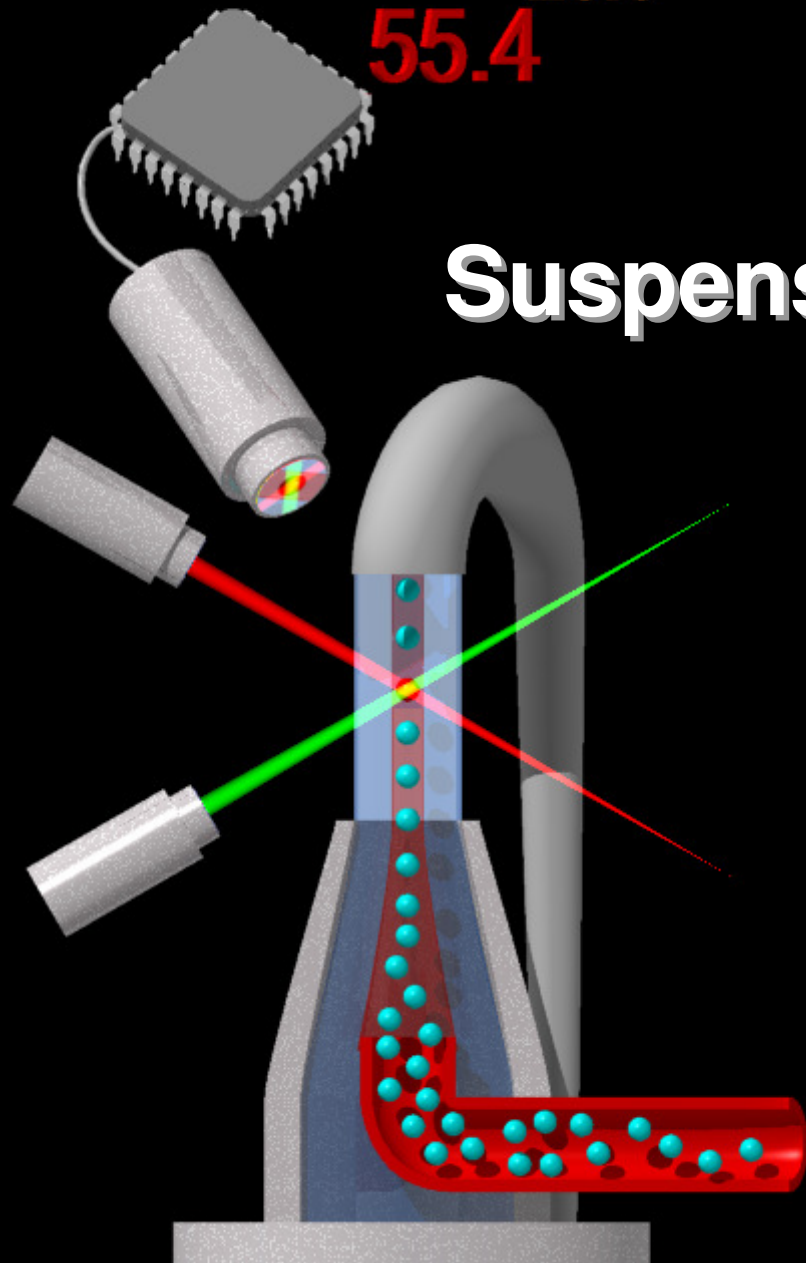
**Multi-Analyte Profiling**

**with**

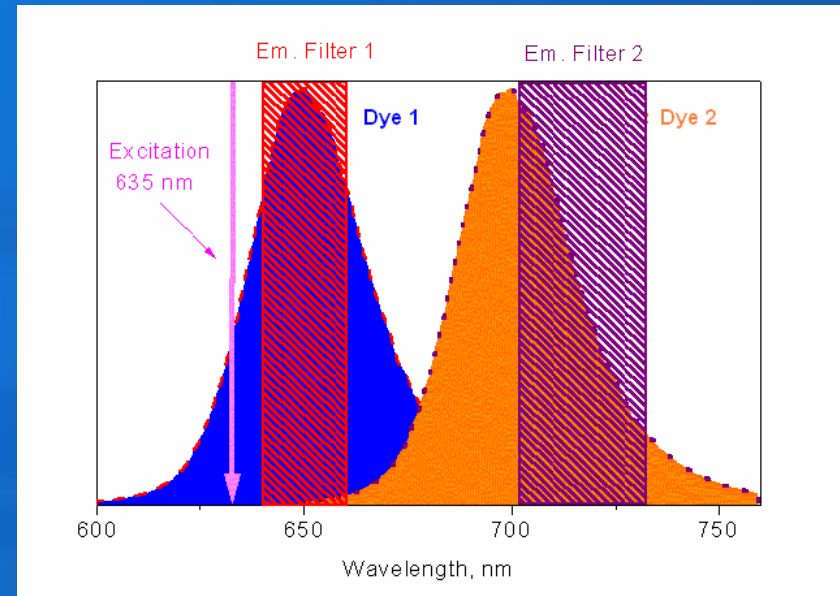
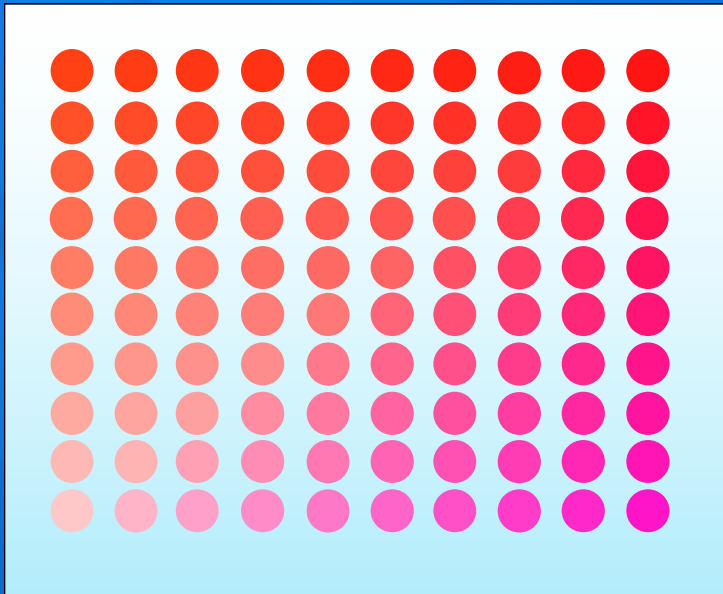
**Suspension Arrays™**

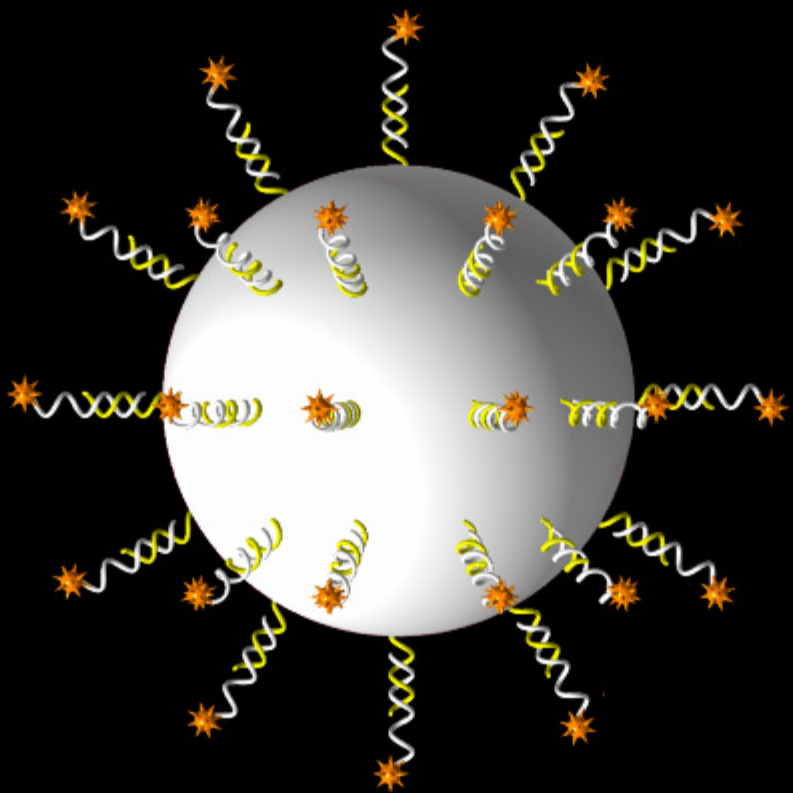
55.4  
25.6  
71.2  
16.8  
42.9  
1000

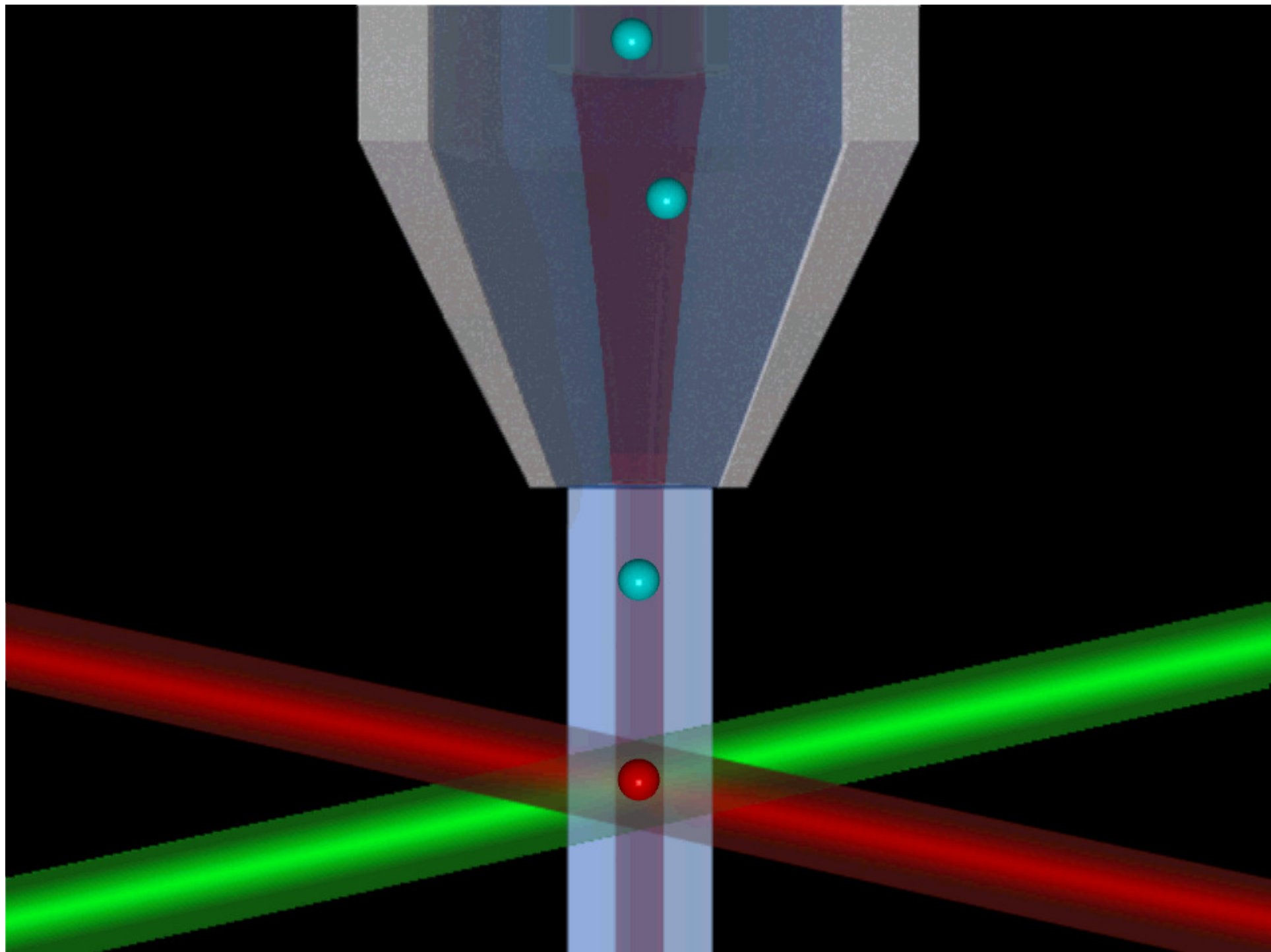
# Suspension Arrays™

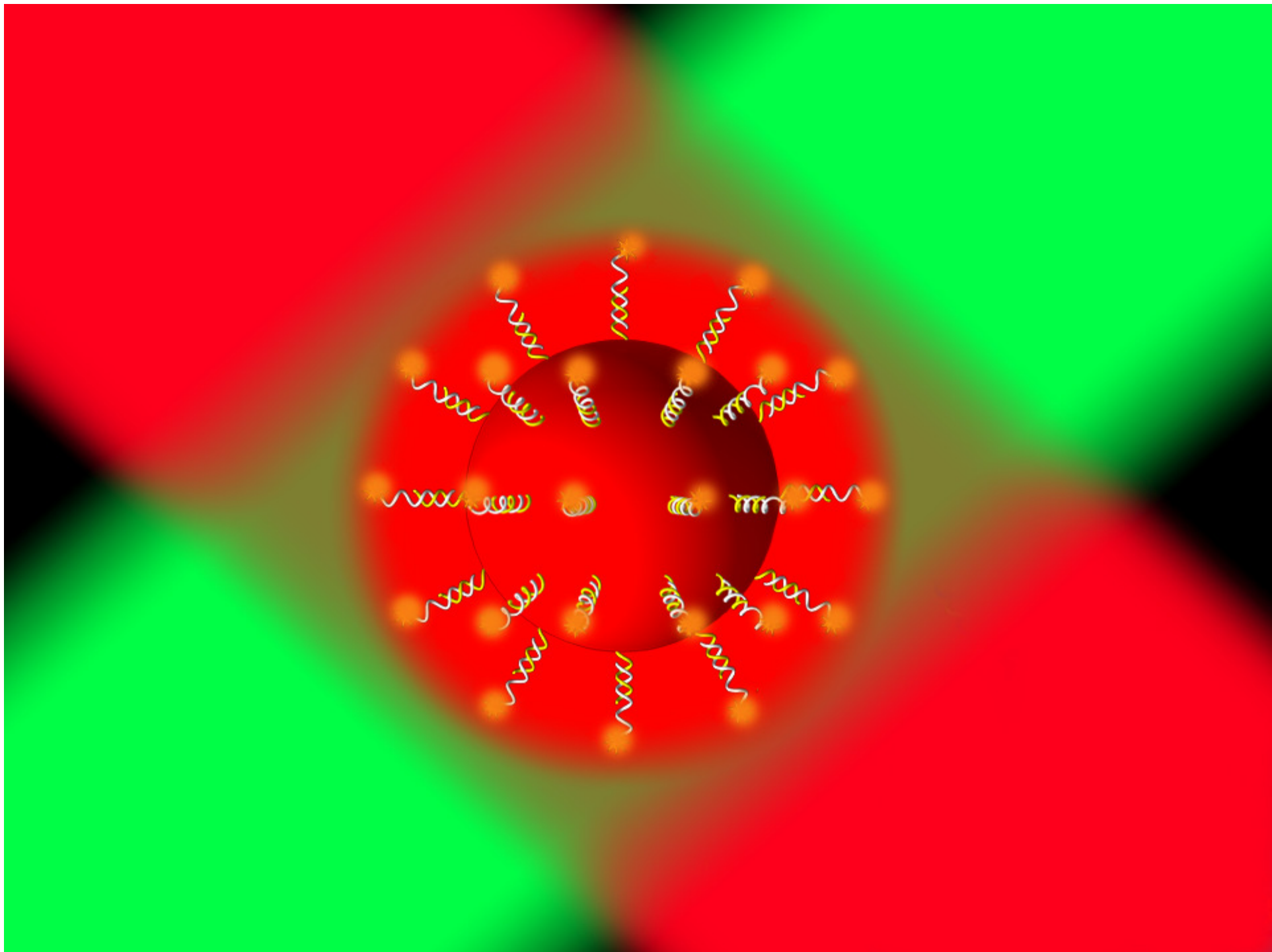


# 100-Plex Bead System

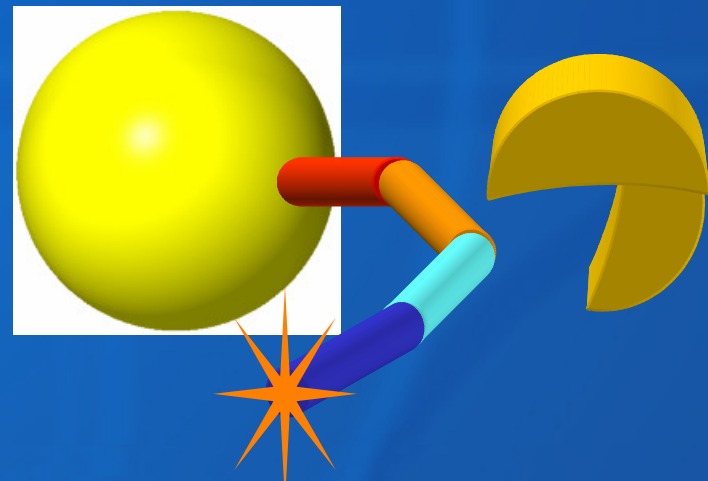
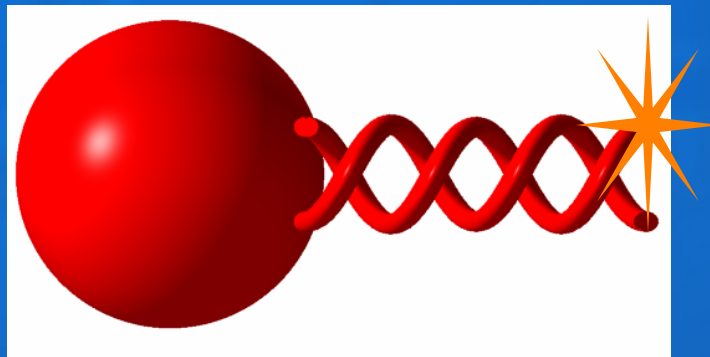
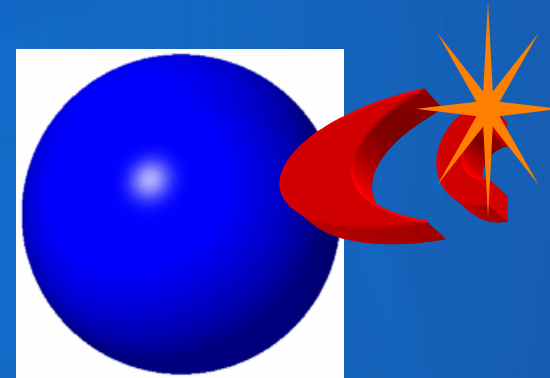
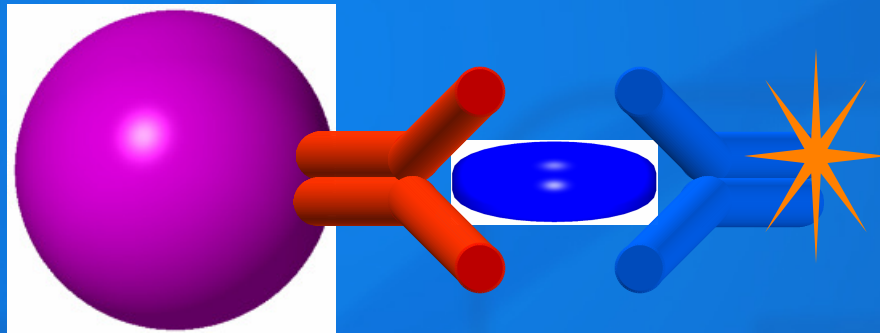




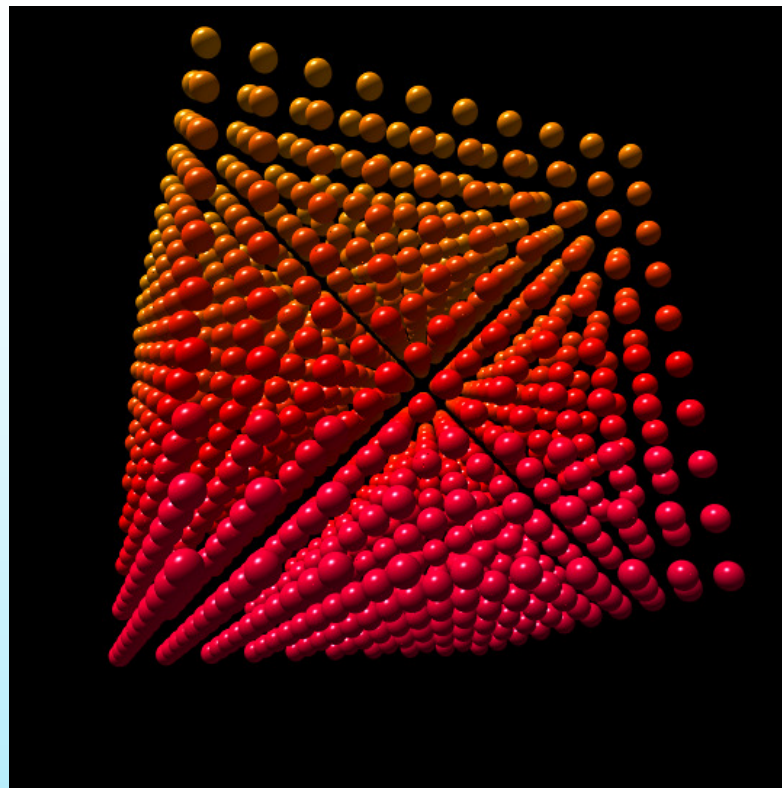
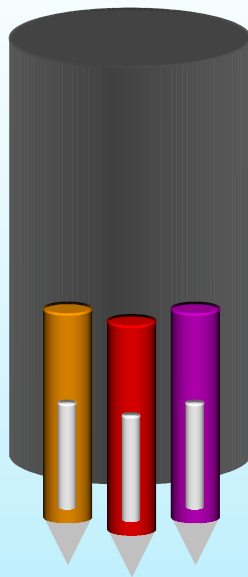




# Applicable Formats



# 1000 Bead Set System





Thank You